

# Artificial Biochemistry

Luca Cardelli

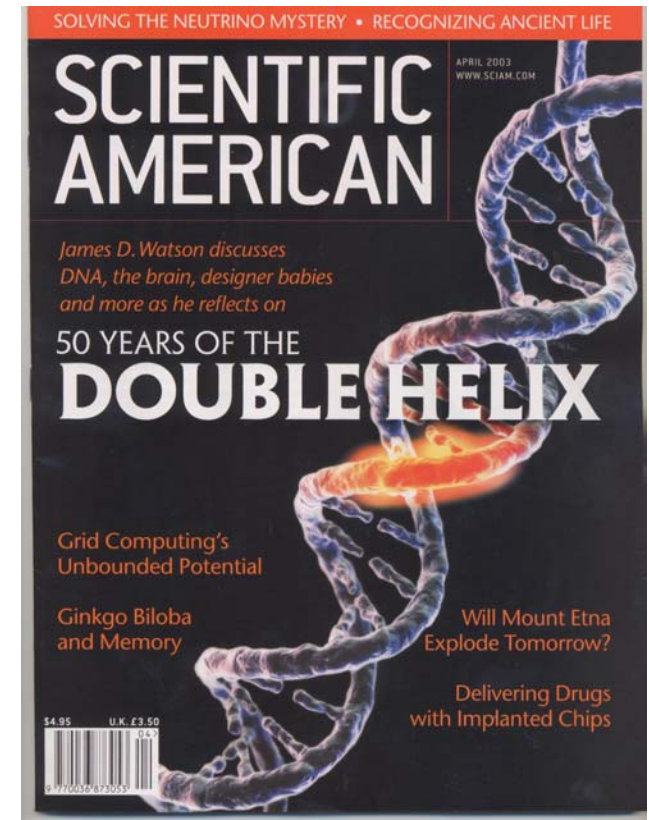
Microsoft Research

LIX Colloquium on Emerging  
Trends in Concurrency Theory  
Paris 2006-11-15

<http://LucaCardelli.name>

# 50 Years of Molecular Cell Biology

- The genome (3.2 GBases) is made of DNA
  - Stores digital information as sequences of 4 different nucleotides
  - Directs protein assembly through RNA and the Genetic Code
- Proteins (1M coded from 25K genes) are made of amino acids
  - Catalyze all biochemical reactions
  - Control metabolism (energy & materials)
  - Process signals, activate genes
- Bootstrapping still a mystery
  - DNA, RNA, proteins, membranes are today interdependent. Not clear who came first
  - Not understood, not essential for us



# Towards Systems Biology

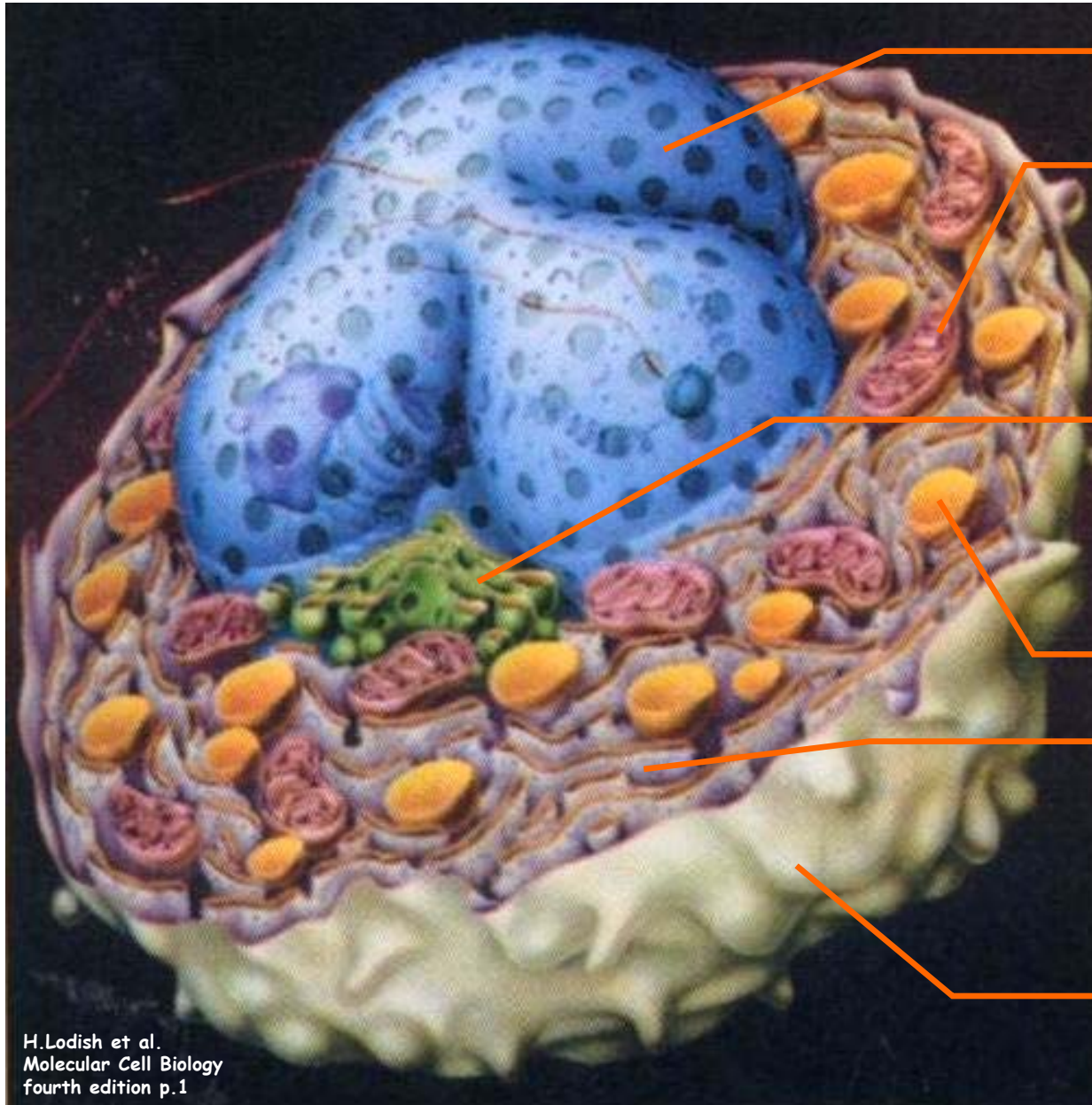
- To reverse-engineer nature
  - You cannot do it one molecule at a time (quickly running out of biologists)
  - You cannot do it one component at a time (without understanding their interactions)
- Understanding how "the systems" works
  - Behavior comes from **complex patterns of interactions between components**
  - Components themselves often irrelevant (analogous function across subsystem/species)
- New(ish) approach (as proposed by biologists)
  - Experimentally: massive data gathering and data mining (e.g. Genome projects)
  - Conceptually: modeling and analyzing large networks (i.e. interactions) of components
- Active research
  - Chemical origin of life and evolutionary processes
  - Systematic mapping of any system of interest
  - Medical control of biological system (no magic bullets)

# Reverse-Engineer This!

## Eukaryotic Cell

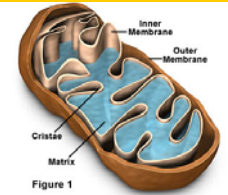
(10~100 trillion in human body)

Membranes everywhere

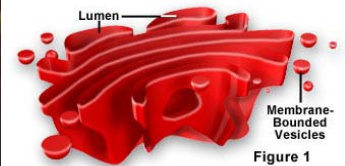


Nuclear membrane

Mitochondria

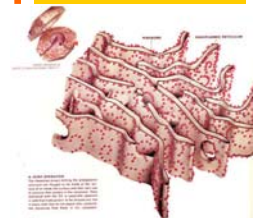


Golgi



Vesicles

E.R.



Plasma membrane (<10% of all membranes)



H.Lodish et al.  
Molecular Cell Biology  
fourth edition p.1

# ...While Modeling It

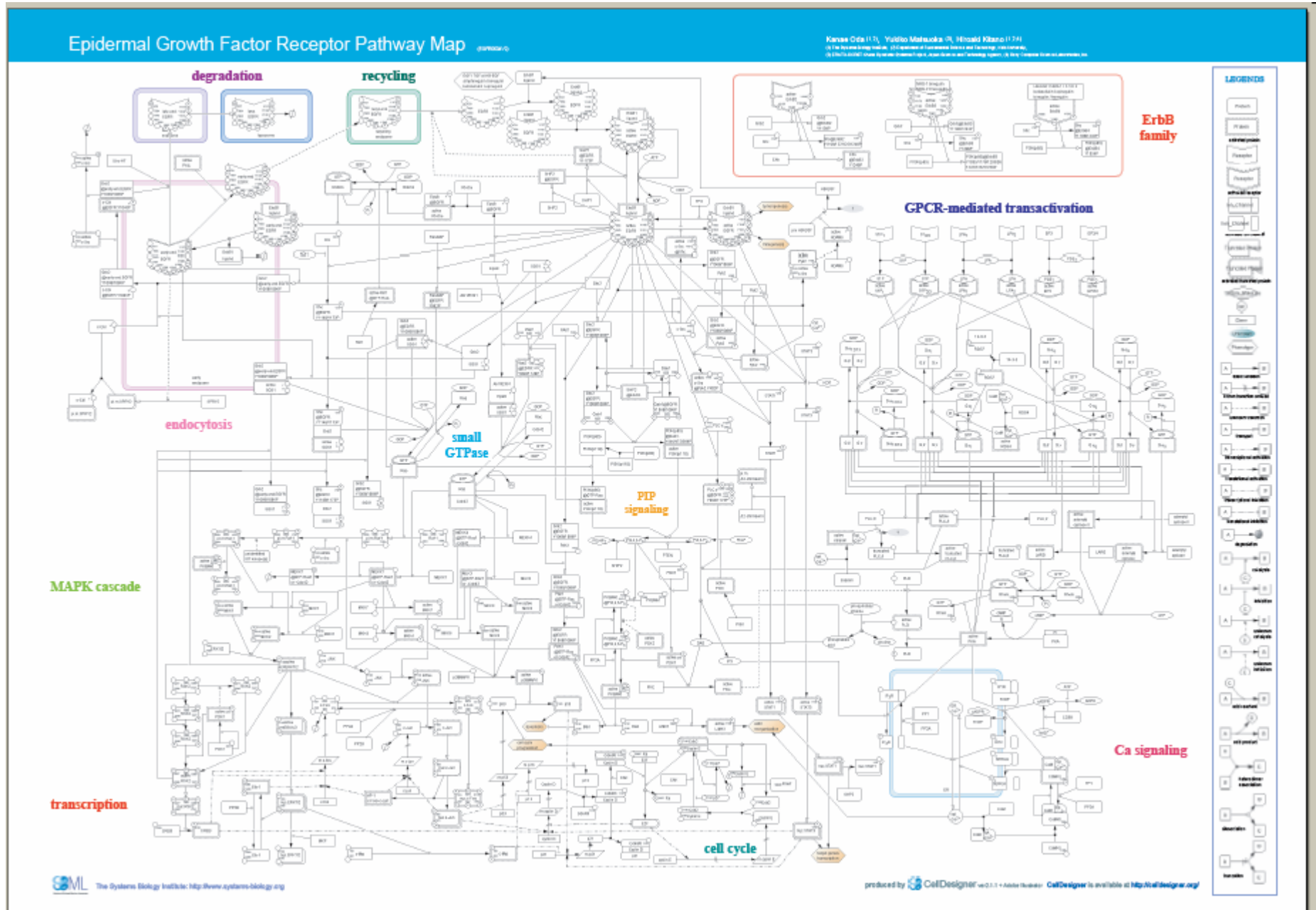
- Even if we understood it, how would we model it?
  - Millions of differential equations? Hmmm...
  - Highly, *highly*, concurrent and asynchronous
  - Stochastic (nondeterministic and discontinuous)
- And we will have to model it in order to understand it.
  - Simulation and analysis are key to understanding interactions
- What's peculiar about these systems?
  - They are huge and complicated
  - They are concurrent and unpredictable
  - There is no documentation
  - You understand them by looking at traces and dumps
  - Doesn't that sound familiar?

# Stochastic Collectives

# Stochastic Collectives

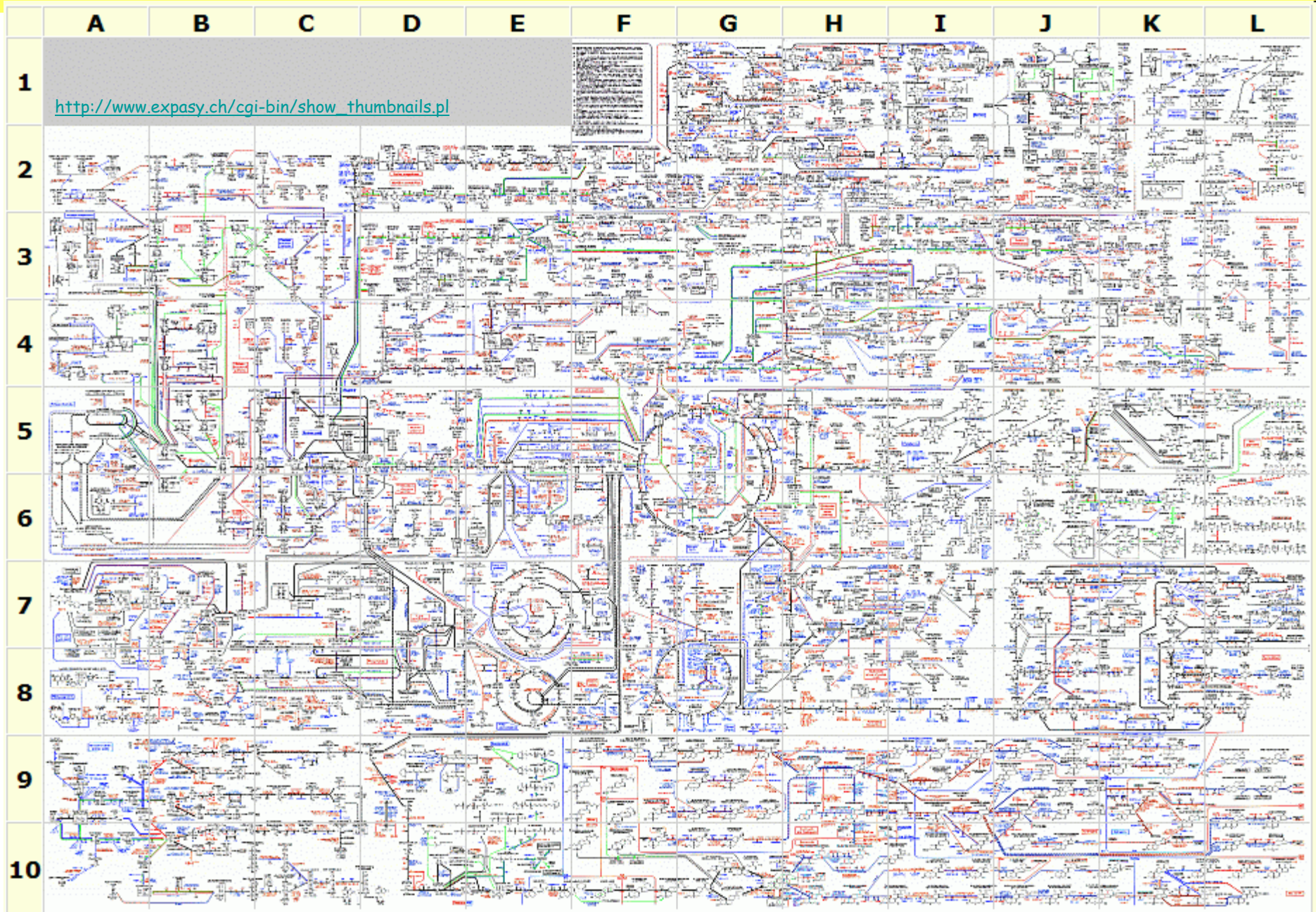
- "Collective":
  - A large set of interacting finite state automata:
    - Not quite language automata ("large set")
    - Not quite cellular automata ("interacting" but not on a grid)
    - Not quite process algebra ("collective behavior")
    - Cf. multi-agent systems and swarm intelligence
- "Stochastic":
  - Interactions have *rates*
    - Not quite discrete (hundreds or thousands of components)
    - Not quite continuous (non-trivial stochastic effects)
    - Not quite hybrid (no "switching" between regimes)
- Very much like biochemistry
  - Which is a large set of stochastically interacting molecules/proteins
  - Are proteins **finite state** and subject to automata-like **transitions**?
    - Let's say they are, at least because:
    - Much of the knowledge being accumulated in Systems Biology is described as state transition diagrams [Kitano].

# State Transitions

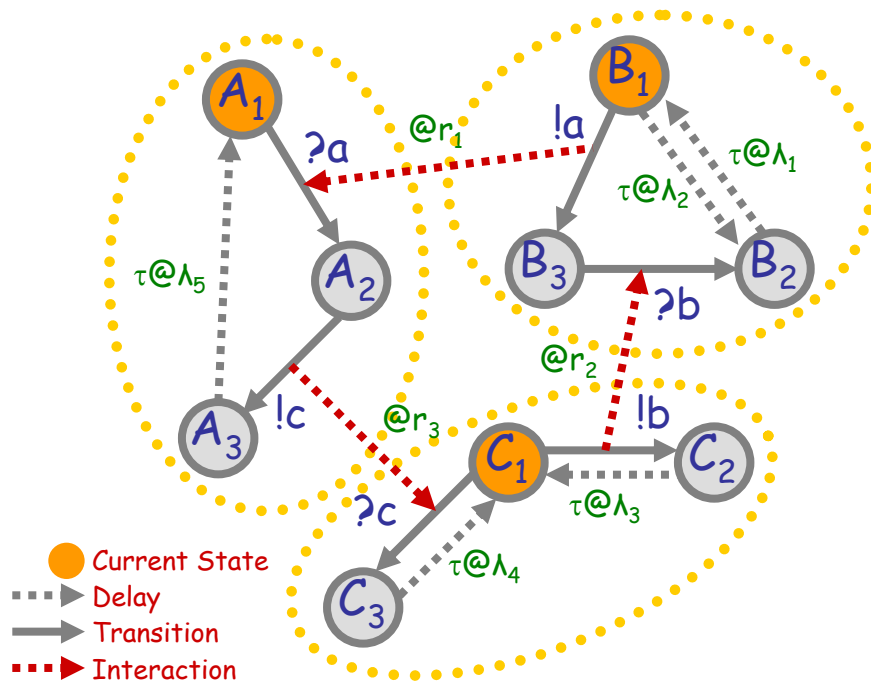




# Compositionality (NOT!)



# Interacting Automata

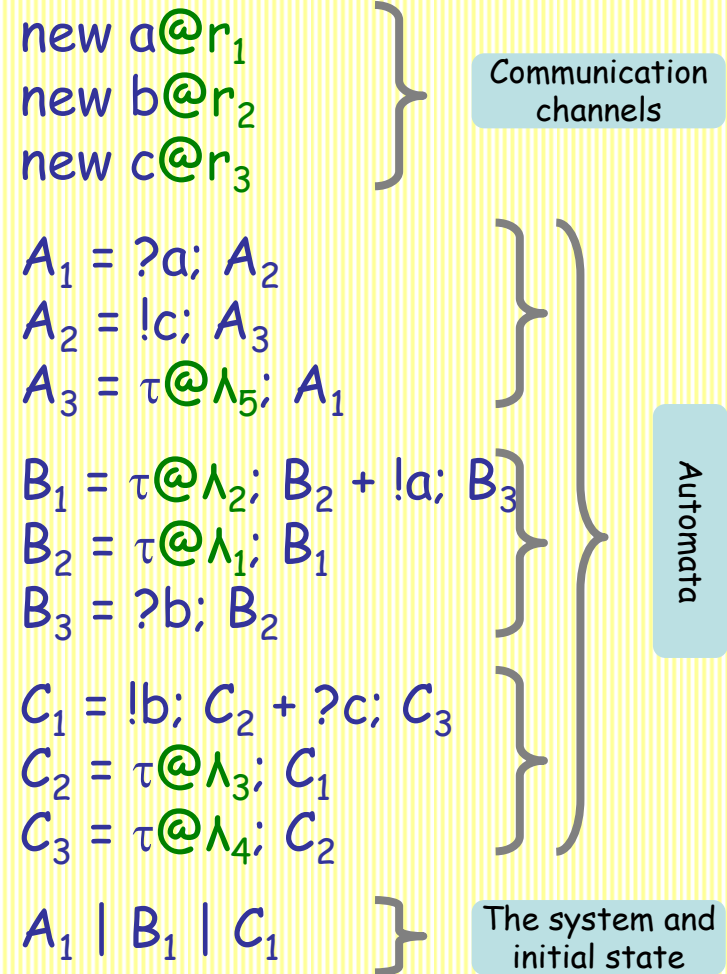


**Communicating automata:** a graphical FSA-like notation for “finite state restriction-free  $\pi$ -calculus processes”. **Interacting automata** do not even exchange values on communication.

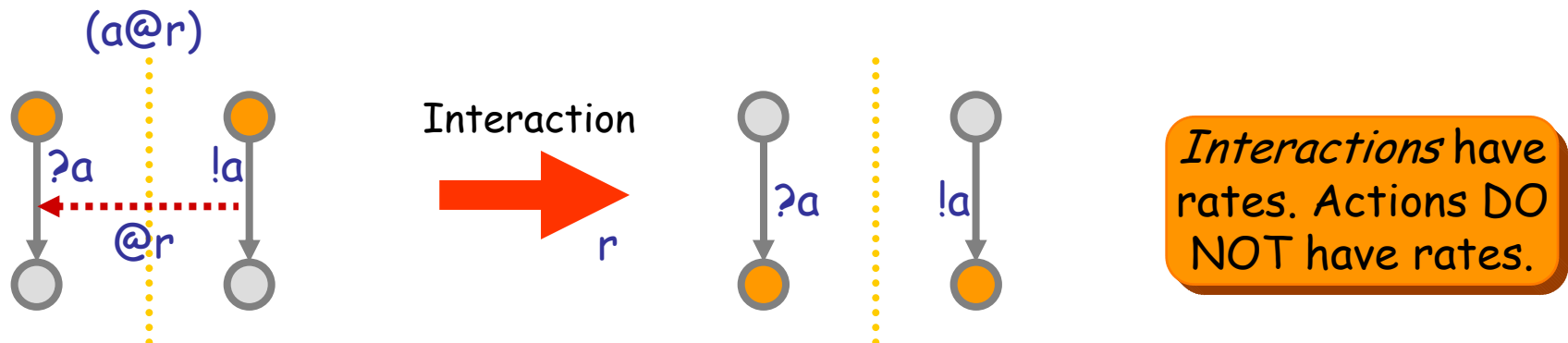
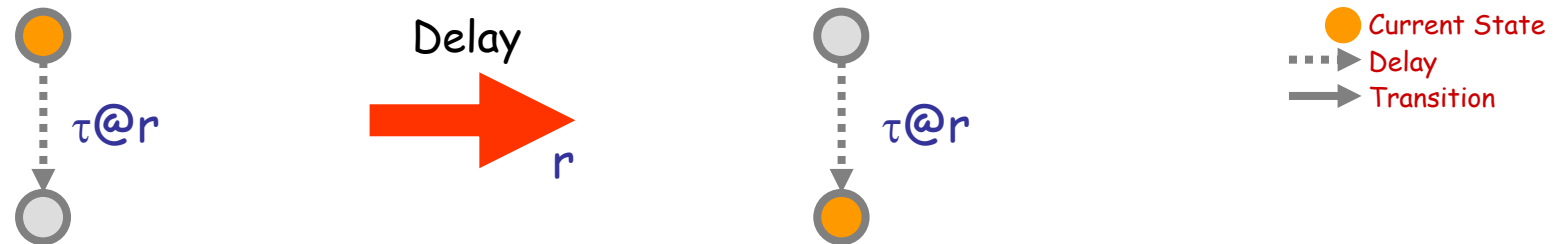
The stochastic version has *rates* on communications, and delays.

“Finite state” means: no composition or restriction inside recursion.

Analyzable by standard Markovian techniques, by first computing the “product automaton” to obtain the underlying finite Markov transition system. [Buchholz]



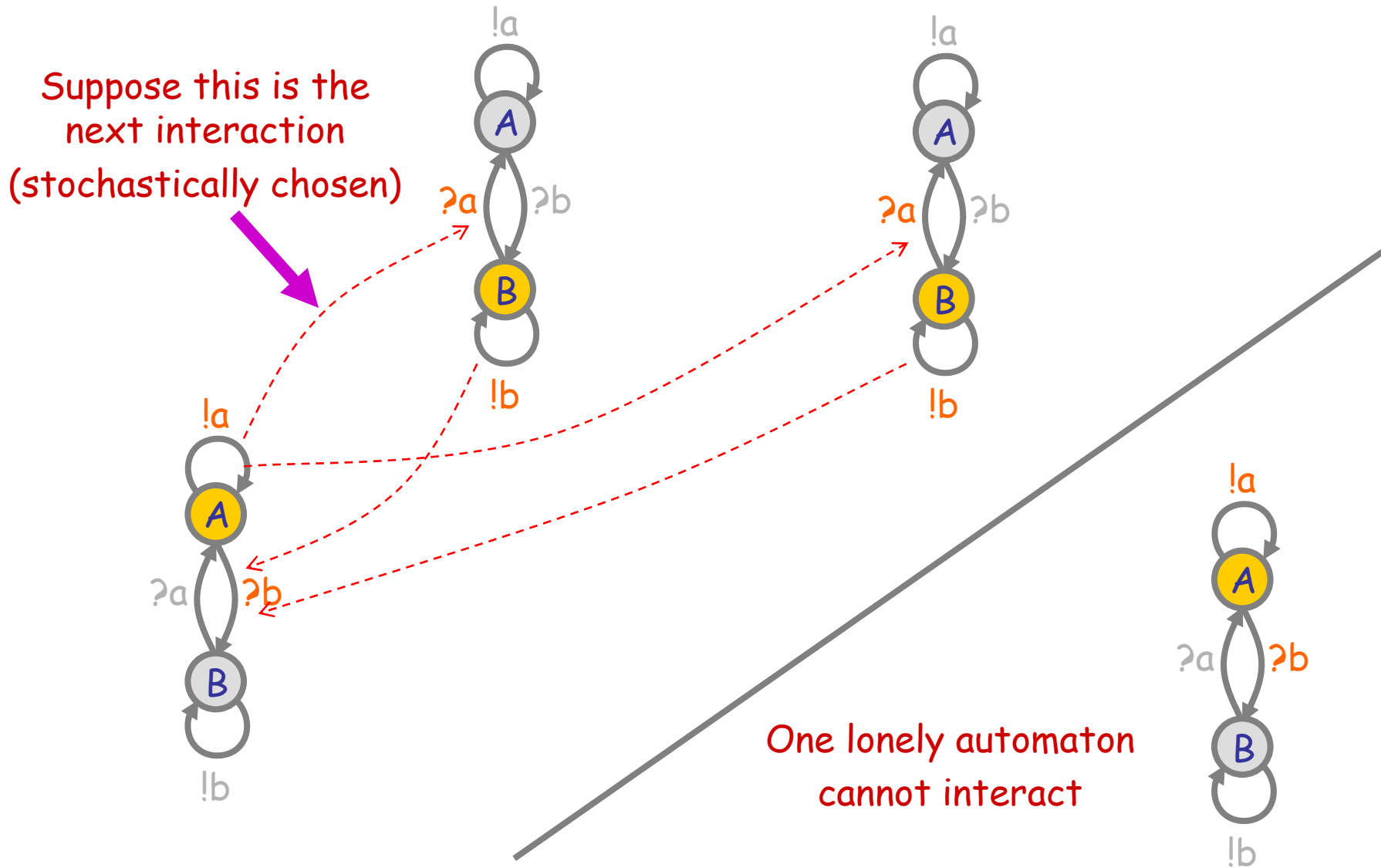
# Interacting Automata Transition Rules



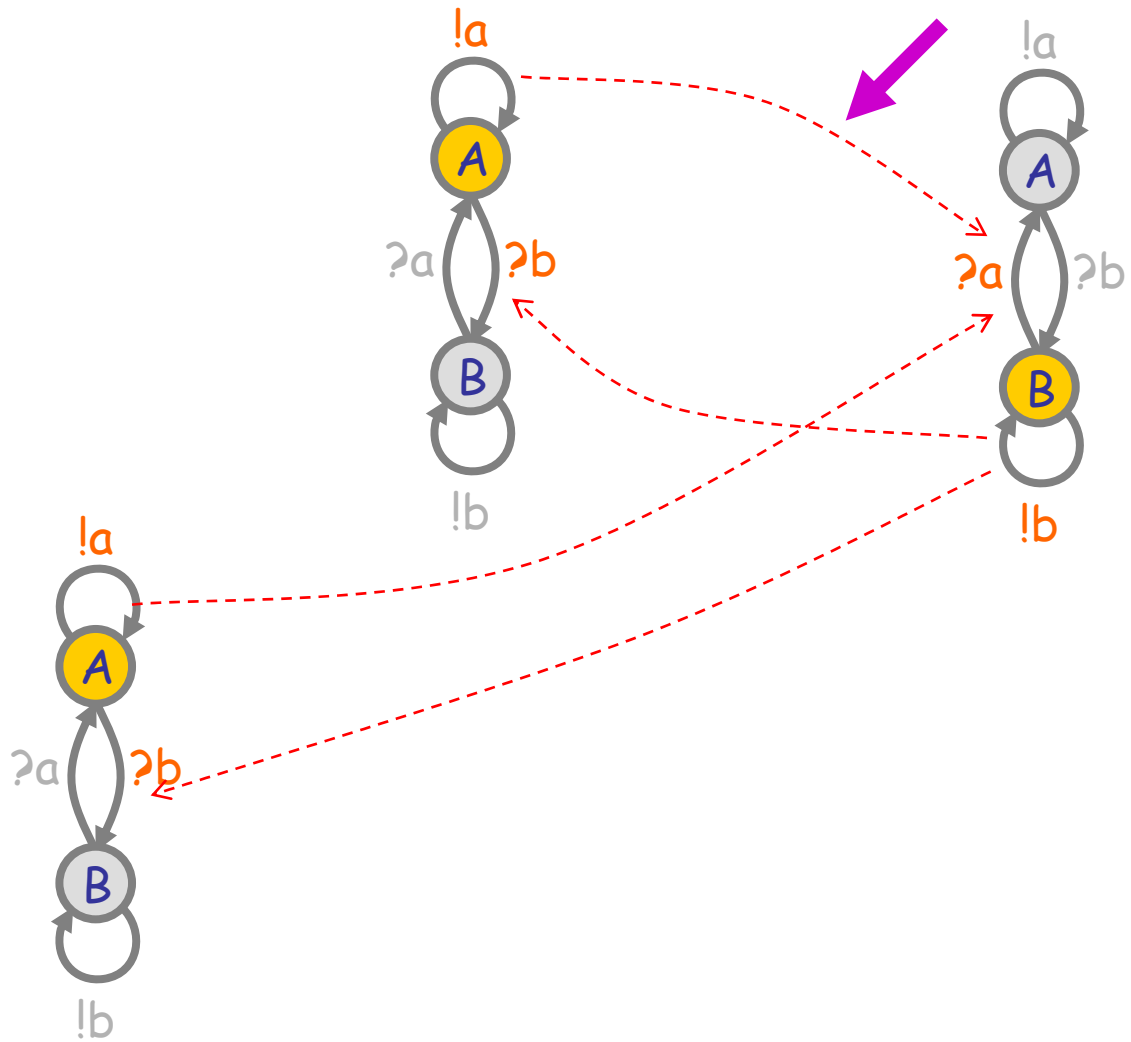
**Q: What kind of mass behavior can this produce?**

(We need to understand that if want to understand biochemical systems.)

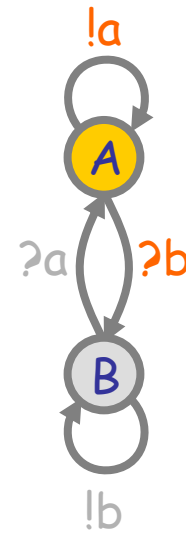
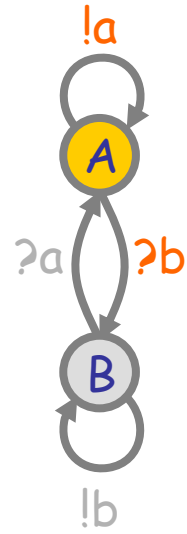
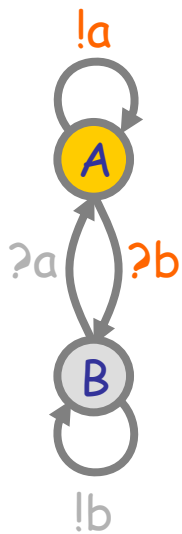
# Interactions in a Population



# Interactions in a Population

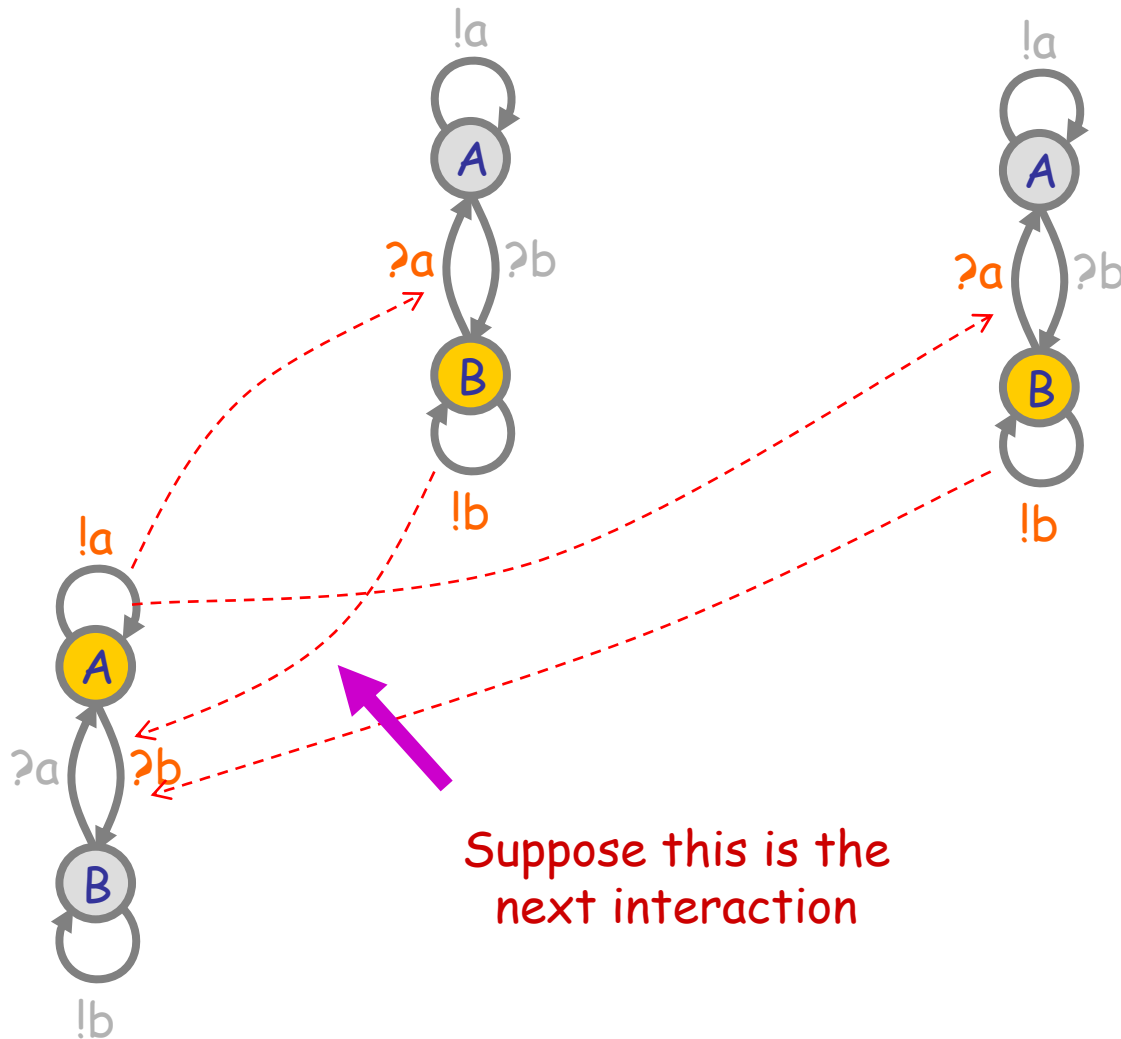


# Interactions in a Population

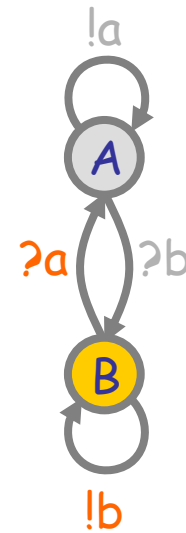
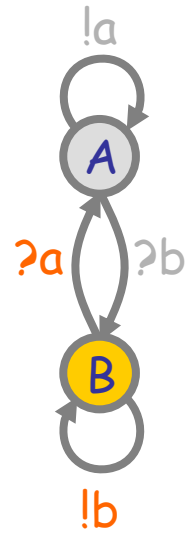
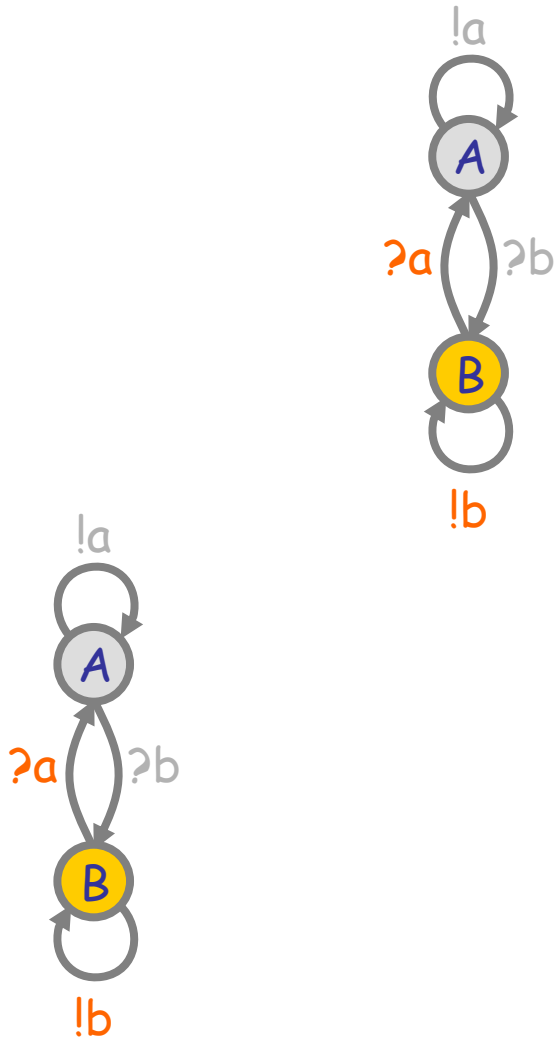


All-A stable  
population

# Interactions in a Population (2)



# Interactions in a Population (2)

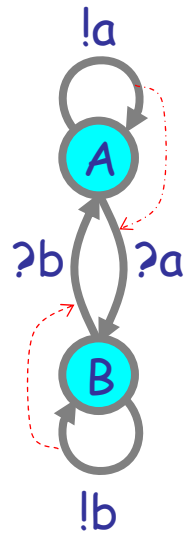


All-B stable population

Nondeterministic population behavior ("multistability")



# Groupies and Celebrities



## Celebrity

(does not want to be like somebody else)

```
directive sample 0.1 200
directive plot A(); B()
```

```
new a@1.0:chan()
new b@1.0:chan()
```

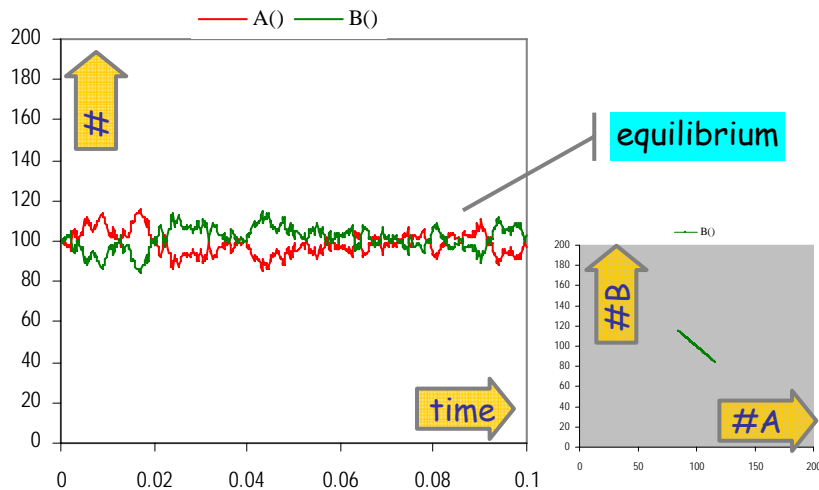
```
let A() = do !a; A() or ?a; B()
and B() = do !b; B() or ?b; A()
```

```
run 100 of (A() | B())
```

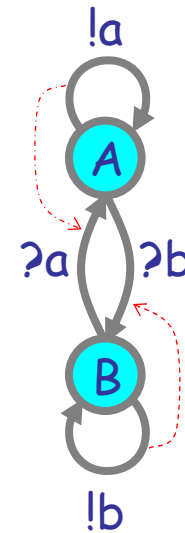
a@1.0

b@1.0

A stochastic collective of celebrities:



Stable because as soon as a A finds itself in the majority, it is more likely to find somebody in the same state, and hence change, so the majority is weakened.



## Groupie

(wants to be like somebody different)

```
directive sample 0.1 200
directive plot A(); B()
```

```
new a@1.0:chan()
new b@1.0:chan()
```

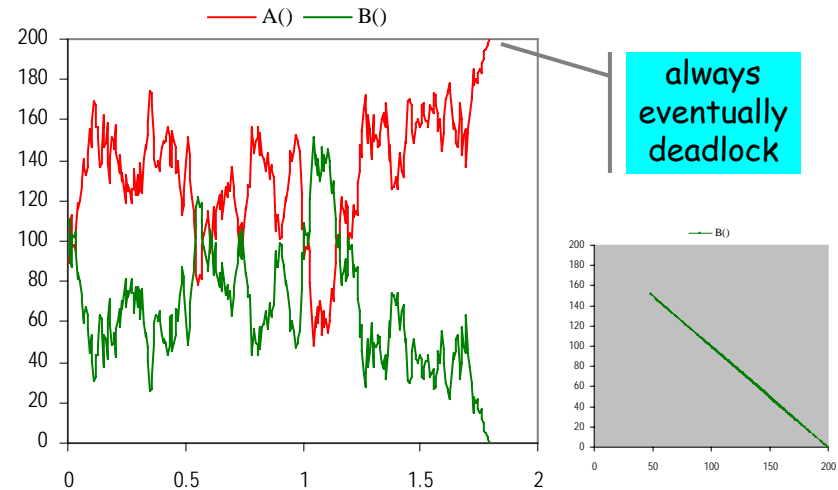
```
let A() = do !a; A() or ?b; B()
and B() = do !b; B() or ?a; A()
```

```
run 100 of (A() | B())
```

a@1.0

b@1.0

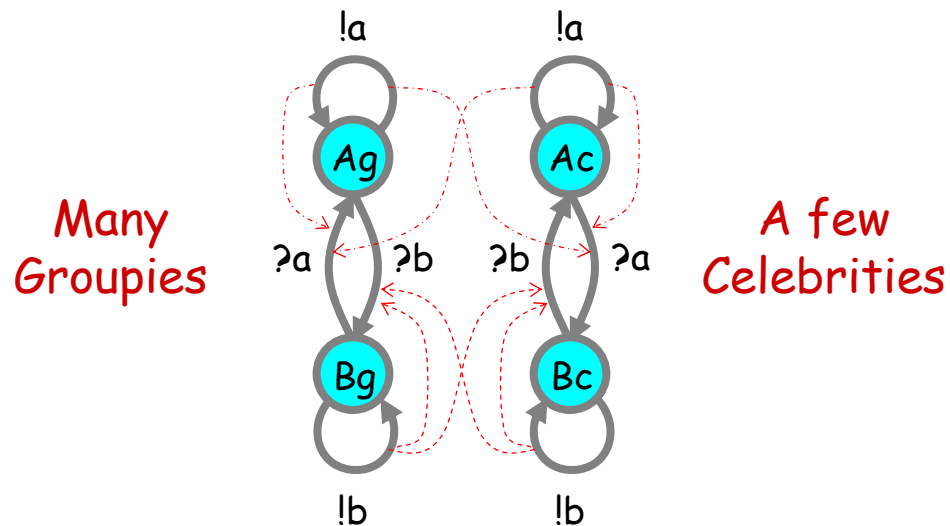
A stochastic collective of groupies:



Unstable because within an A majority, an A has difficulty finding a B to emulate, but the few B's have plenty of A's to emulate, so the majority may switch to B. Leads to deadlock when everybody is in the same state and there is nobody different to emulate.

# Both Together

A way to break the deadlocks: Groupies with just a few Celebrities



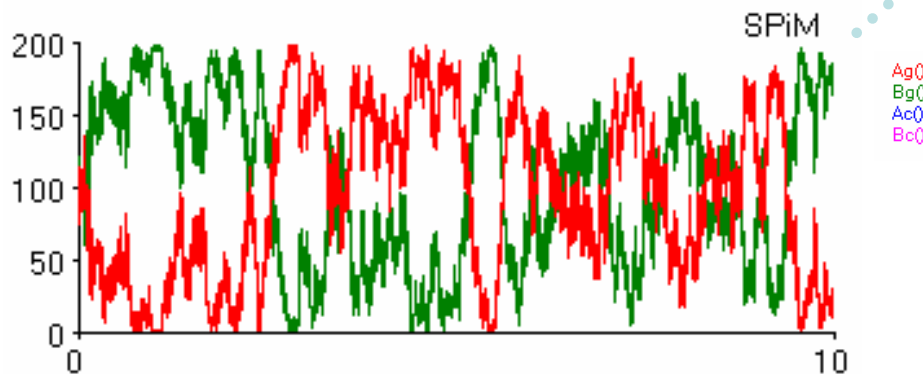
```
directive sample 10.0
directive plot Ag(); Bg(); Ac(); Bc()

new a@1.0:chan()
new b@1.0:chan()

let Ac() = do !a; Ac() or ?a; Bc()
and Bc() = do !b; Bc() or ?b; Ac()

let Ag() = do !a; Ag() or ?b; Bg()
and Bg() = do !b; Bg() or ?a; Ag()

run 1 of Ac()
run 100 of (Ag() | Bg())
```



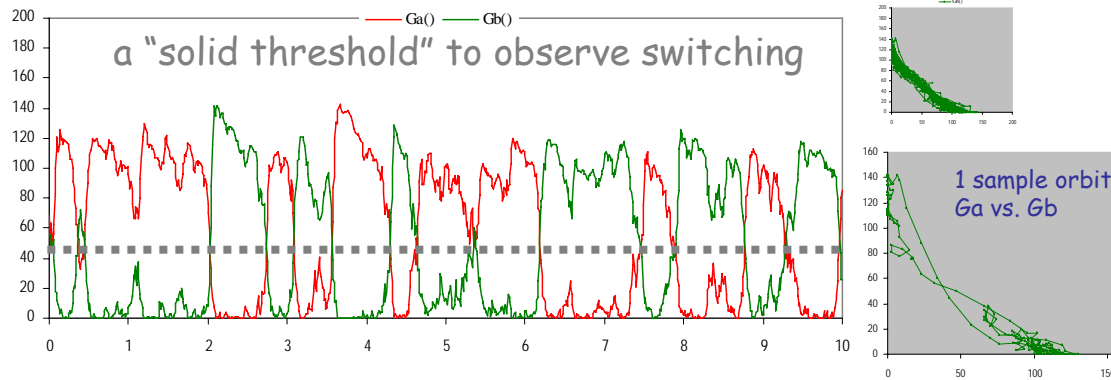
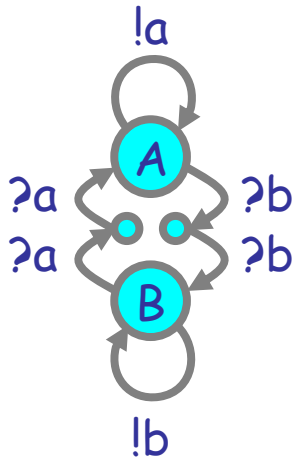
never  
deadlock

A tiny bit of  
"noise" can make a  
huge difference

Regularity can arise not far from chaos

# Hysteric Groupies

We can get more regular behavior from groupies if they "need more convincing", or "hysteresis" (history-dependence), to switch states.



```
directive sample 10.0 1000
directive plot Ga(); Gb()

new a@1.0:chan()
new b@1.0:chan()

let Ga() = do !a; Ga() or ?b; ?b; Gb()
and Gb() = do !b; Gb() or ?a; ?a; Ga()

let Da() = !a; Da()
and Db() = !b; Db()

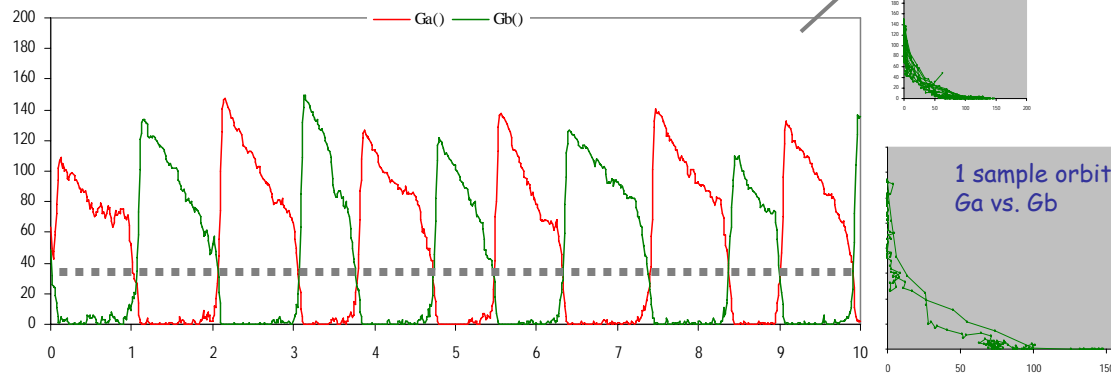
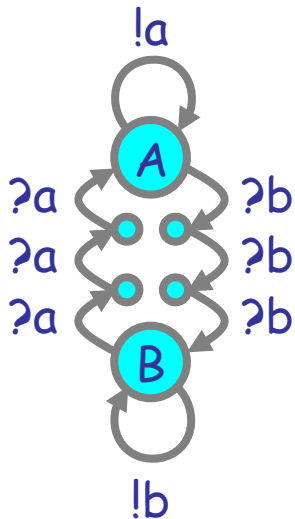
run 100 of (Ga() | Gb())
run 1 of (Da() | Db())
```



(With doping to break deadlocks)

N.B.: It will not oscillate without doping (noise)

"regular" oscillation



```
directive sample 10.0 1000
directive plot Ga(); Gb()

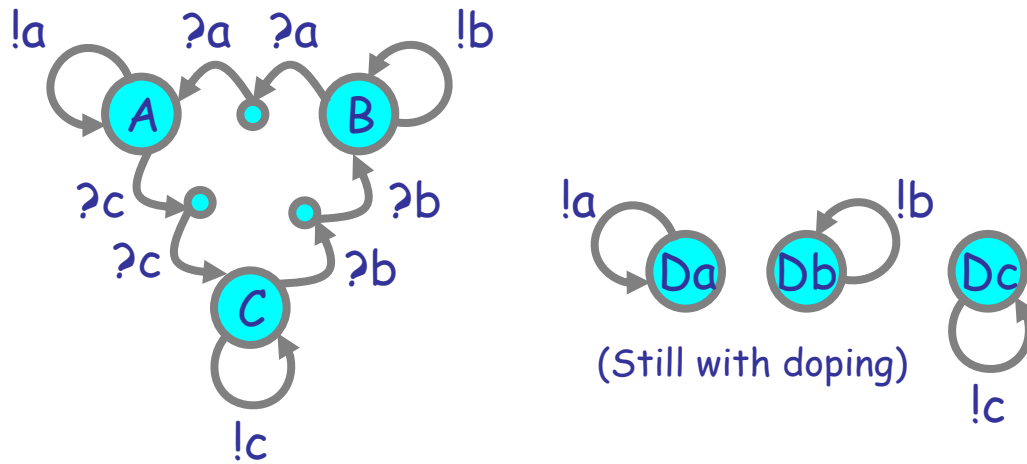
new a@1.0:chan()
new b@1.0:chan()

let Ga() = do !a; Ga() or ?b; ?b; ?b; Gb()
and Gb() = do !b; Gb() or ?a; ?a; ?a; Ga()

let Da() = !a; Da()
and Db() = !b; Db()

run 100 of (Ga() | Gb())
run 1 of (Da() | Db())
```

# Hysteric 3-Way Groupies



```
directive sample 3.0 1000
directive plot A(); B(); C()
```

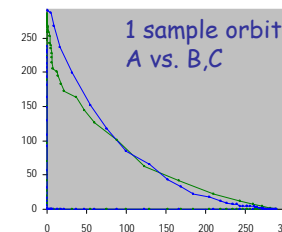
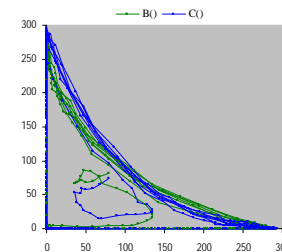
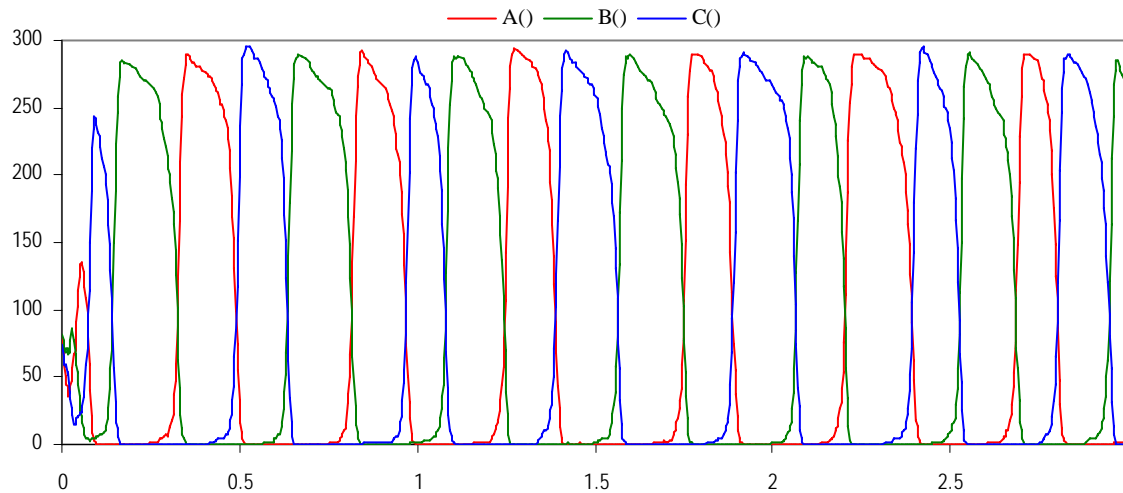
```
new a@1.0:chan()
new b@1.0:chan()
new c@1.0:chan()
```

```
let A() = do !a; A() or ?c; ?c; C()
and B() = do !b; B() or ?a; ?a; A()
and C() = do !c; C() or ?b; ?b; B()
```

```
let Da() = !a; Da()
and Db() = !b; Db()
and Dc() = !c; Dc()
```

```
run 100 of (A() | B() | C())
run 1 of (Da() | Db() | Dc())
```

N.B.: It will not oscillate without doping (noise)



# Semantics of Collective Behavior

# "Micromodels": Continuous Time Markov Chains

- The underlying semantics of stochastic  $\pi$ -calculus (and stochastic interacting automata). Well established in many ways.
  - Automata with rates on transitions.
- "The" correct semantics for chemistry, executable.
  - Gillespie stochastic simulation algorithm
- Lots of advantages
  - Compositional, compact, mechanistic, etc.
- But do not give a good sense of "collective" properties.
  - Yes one can do simulation.
  - Yes one can do program analysis.
  - Yes one can do modelchecking.
  - But somewhat lacking in "analytical properties" and "predictive power".

# "Macromodels": Ordinary Differential Equations

- The classical semantics of collective behavior.
  - E.g. kinetic theory of gasses.
  - They always ask: "How does your automata model relate to the 75 ODE models in the literature?"
- Going from processes/automata to ODEs directly:
  - *In principle*: just write down the **Rate Equation**: [Calder, Hillston]
    - Let  $[S]$  be the "number of processes in state  $S$ " as a function of time.
    - Define for each state  $S$ :  
 $[S]' =$  (rate of change of the number of processes in state  $S$ )  
Cumulative rate of transitions from any state  $S'$  to state  $S$ , times  $[S']$ ,  
minus cumulative rate of transitions from  $S$  to any state  $S''$ , times  $[S]$ .
  - Intuitive (rate = inflow minus outflow), but clumsy to write down precisely.
- Going to ODEs indirectly through chemistry
  - If we first convert processes to chemical reactions, then we can convert to ODEs by standard means!



# From Chemistry to ODEs



# Chemical Reactions

$A \rightarrow^r B_1 + \dots + B_n$	Degradation	$[A]^{\bullet} = -r[A]$	Exponential Decay
$A_1 + A_2 \rightarrow^r B_1 + \dots + B_n$	Asymmetric Collision	$[A_i]^{\bullet} = -r[A_1][A_2]$	Mass Action Law
$A + A \rightarrow^r B_1 + \dots + B_n$	Symmetric Collision	$[A]^{\bullet} = -r[A]([A]-1)$	Mass Action Law

(assuming  $A \neq B_i \neq A_j$  for all  $i, j$ )

No other reactions!

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## The chemical Langevin equation

Daniel T. Gillespie<sup>a)</sup>  
 Research Department, Code 4T4100D, Naval Air Warfare Center, China Lake, California 93555

Genuinely *trimolecular* reactions do not physically occur in dilute fluids with any appreciable frequency. *Apparently* trimolecular reactions in a fluid are usually the combined result of two bimolecular reactions and one monomolecular reaction, and involve an additional short-lived species.

## Chapter IV: Chemical Kinetics

[David A. Reckhow, CEE 572 Course]

... reactions may be either elementary or non-elementary. Elementary reactions are those reactions that occur exactly as they are written, without any intermediate steps. These reactions **almost always involve just one or two reactants**. ... Non-elementary reactions involve a series of two or more elementary reactions. Many complex environmental reactions are non-elementary. In general, **reactions with an overall reaction order greater than two, or reactions with some non-integer reaction order are non-elementary**.

## THE COLLISION THEORY OF REACTION RATES

[www.chemguide.co.uk](http://www.chemguide.co.uk)

The chances of all this happening if your reaction needed a collision involving more than 2 particles are remote. All three (or more) particles would have to arrive at exactly the same point in space at the same time, with everything lined up exactly right, and having enough energy to react. That's not likely to happen very often!

Trimolecular reactions:



the measured "r" is an (imperfect) aggregate of e.g.:



Enzymatic reactions:

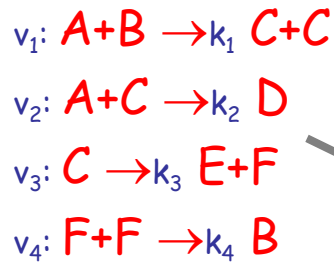


the "r" is given by Michaelis-Menten: (approximated steady-state) laws:



*Reactions have rates. Molecules do not have rates.*

# From Reactions to ODEs

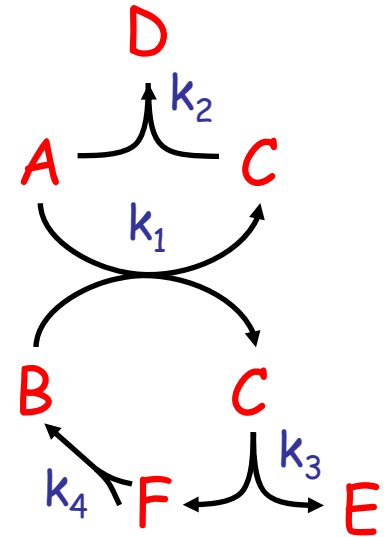


Write the coefficients by columns

		reactions			
species	N	$v_1$	$v_2$	$v_3$	$v_4$
	A	-1	-1		
	B	-1			1
	C	2	-1	-1	
	D		1		
	E			1	
	F			1	-2

**X**

Stoichiometric Matrix



CAVEAT: A deterministic approximation of a stochastic system (i.e. possibly misleading)

Quantity changes

Stoichiometric matrix

Rate laws

$$[X]^\bullet = N \cdot I$$

Read the concentration changes from the rows

$$\begin{aligned}
 [A]^\bullet &= -I_1 - I_2 \\
 [B]^\bullet &= -I_1 + I_4 \\
 [C]^\bullet &= 2I_1 - I_2 - I_3 \\
 [D]^\bullet &= I_2 \\
 [E]^\bullet &= I_3 \\
 [F]^\bullet &= I_3 - 2I_4
 \end{aligned}$$

E.g.  $[A]^\bullet = -k_1[A][B] - k_2[A][C]$

Set a rate law for each reaction (Degradation/Asymmetric/Symmetric)

I	
$I_1$	$k_1[A][B]$
$I_2$	$k_2[A][C]$
$I_3$	$k_3[C]$
$I_4$	$k_4[F]([F]-1)/2$

**X**: chemical species  
**[-]**: quantity of molecules  
**I**: rate laws  
**k**: kinetic parameters  
**N**: stoichiometric matrix

# From Processes to Chemistry

# Chemical Ground Form (CGF)

$E ::= X_1=M_1, \dots, X_n=M_n$

Definitions ( $n \geq 0$ )

$M ::= \pi_1;P_1 \oplus \dots \oplus \pi_n;P_n$

Molecules ( $n \geq 0$ )

$P ::= X_1 \mid \dots \mid X_n$

Solutions ( $n \geq 0$ )

$\pi ::= \tau_r \ ?n_{(r)} \ !n_{(r)}$

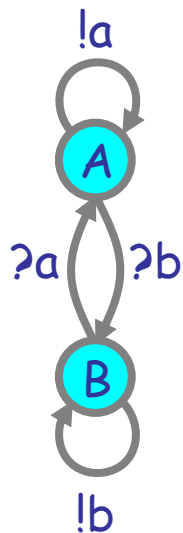
Interactions (delay, input, output)

CGF ::= E,P

Definitions with Initial Conditions

(To translate chemistry back to processes we need a bit more than simple automata: we may have "+" on the right of  $\rightarrow$ , that is we may need "|" after  $\pi$ .)

$\oplus$  is stochastic choice (vs. + for chemical reactions)  
 0 is the null solution ( $P \mid 0 = 0 \mid P = P$ )  
 and null molecule ( $M \oplus 0 = 0 \oplus M = M$ ) ( $\tau_0;P = 0$ )  
 $X_i$  are distinct in E  
 Each name n is assigned a fixed rate r:  $n_{(r)}$



Ex: interacting automata

(which are finite-control CGFs: use "|" only in initial conditions):

$A = !a;A \oplus ?b;B$

Automaton in state A

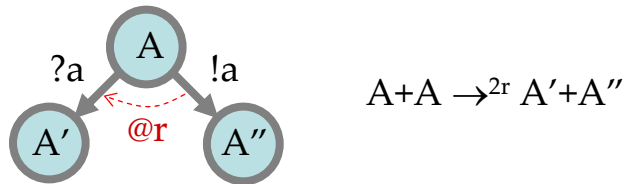
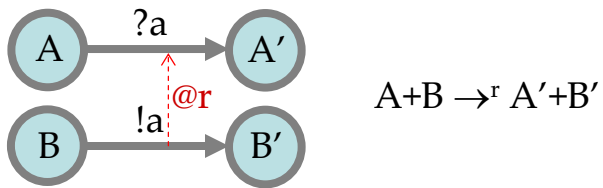
$B = !b;B \oplus ?a;A$

Automaton in state B

$A \mid A \mid B \mid B$

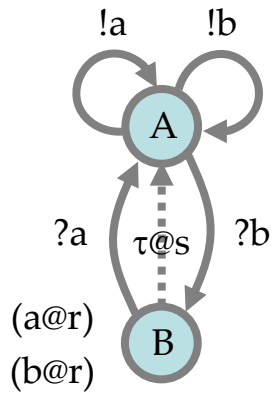
Initial conditions:  
2A and 2B

# Automata (or CGF) to Chemistry



# Process Rate Semantics

# Same Chemistry

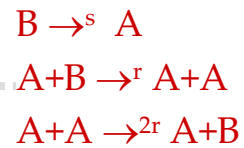
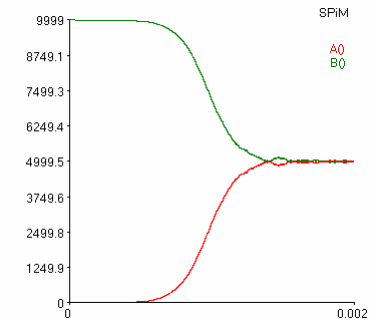


```
directive sample 0.002 10000
directive plot A(); B()

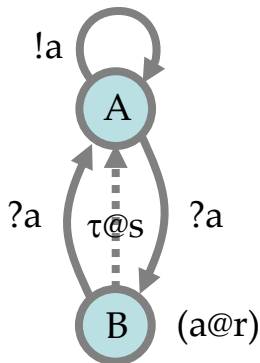
new a@1.0:chan()
new b@1.0:chan()

let A() = do !a; A() or !b; A() or ?b; B()
and B() = do delay@1.0; A() or ?a; A()

run 10000 of B()
```



Same chemistry, hence equivalent automata

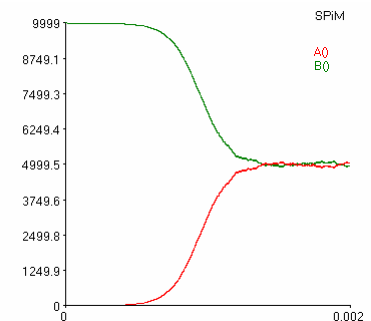


```
directive sample 0.002 10000
directive plot A(); B()

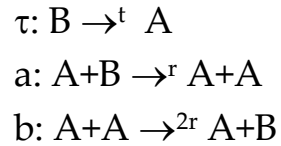
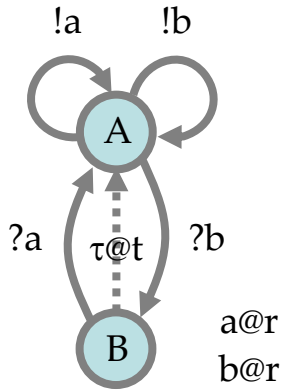
new a@1.0:chan()

let A() = do !a; A() or ?a; B()
and B() = do delay@1.0; A() or ?a; A()

run 10000 of B()
```



# Same ODEs

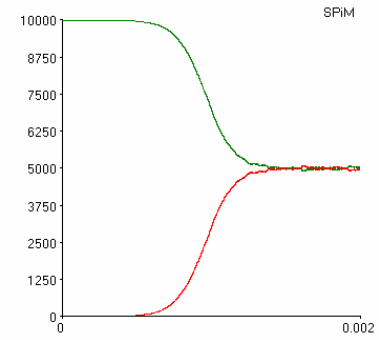


```
directive sample 0.002 10000
directive plot A(); B()

new a@1.0:chan()
new b@1.0:chan()

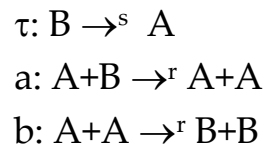
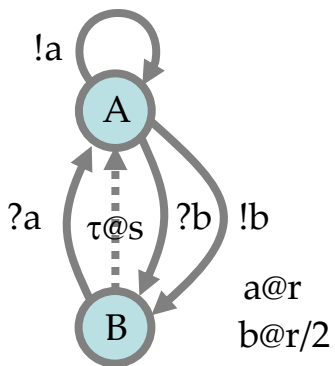
let A() = do !a; A() or !b; A() or ?b; B()
and B() = do delay@1.0; A() or ?a; A()

run 10000 of B()
```



$$\begin{aligned} [A]^* &= t[B] + r[A][B] - r[A]([A]-1) \\ [B]^* &= -t[B] - r[A][B] + r[A]([A]-1) \end{aligned}$$

Different chemistry  
but same ODEs, hence  
equivalent automata

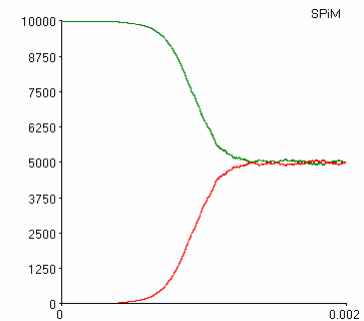


```
directive sample 0.002 10000
directive plot A(); B()

new a@1.0:chan()
new b@0.5:chan()

let A() = do !a; A() or !b; B() or ?b; B()
and B() = do delay@1.0; A() or ?a; A()

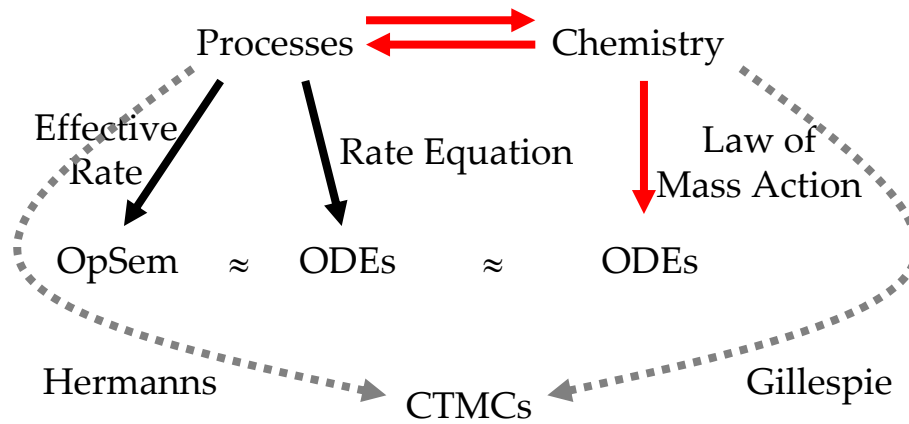
run 10000 of B()
```



$$\begin{aligned} [A]^* &= t[B] + r[A][B] - r[A]([A]-1) \\ [B]^* &= -t[B] - r[A][B] + r[A]([A]-1) \end{aligned}$$



# Semantic Relationships



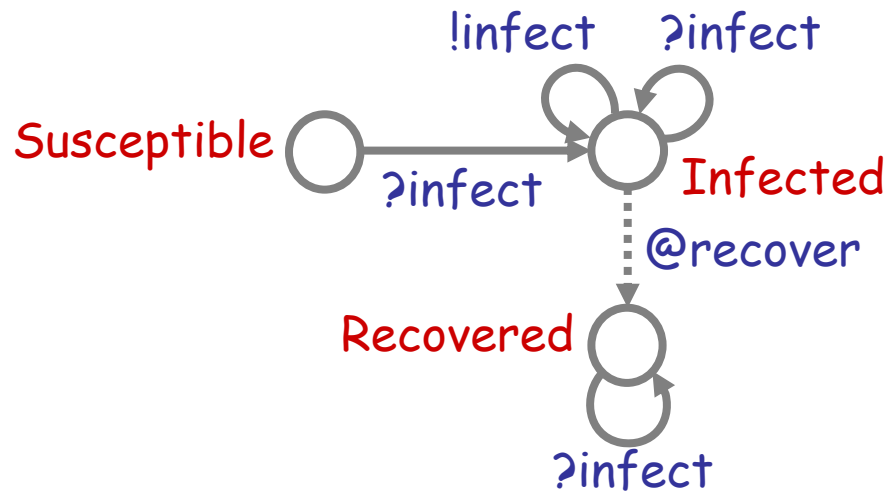


# Epidemics

Kermack, W. O. and McKendrick, A. G. "A Contribution to the Mathematical Theory of Epidemics." *Proc. Roy. Soc. Lond. A* 115, 700-721, 1927.

<http://mathworld.wolfram.com/Kermack-McKendrickModel.html>

# Epidemics



```

directive sample 500.0 1000
directive plot Recovered(); Susceptible(); Infected()

new infect @0.001:chan()
val recover = 0.03

let Recovered() =
  ?infect; Recovered()

and Susceptible() =
  ?infect; Infected()

and Infected() =
  do !infect; Infected()
  or ?infect; Infected()
  or delay@recover; Recovered()

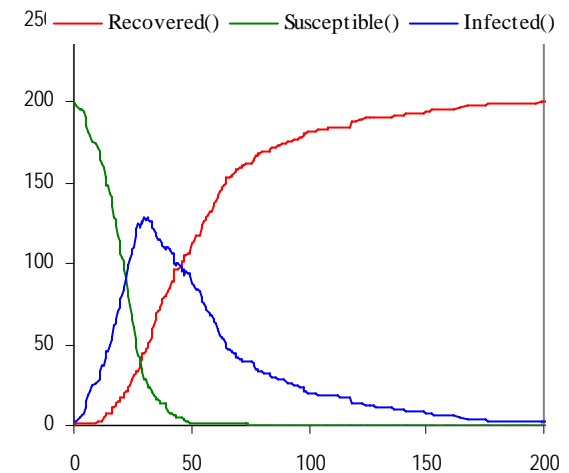
run (200 of Susceptible() | 2 of Infected())
  
```

## Developing the Use of Process Algebra in the Derivation and Analysis of Mathematical Models of Infectious Disease

R. Norman and C. Shankland

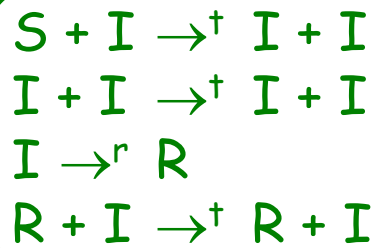
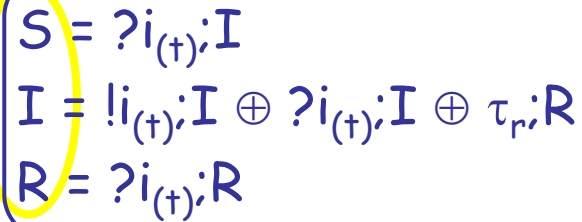
Department of Computing Science and Mathematics, University of Stirling, UK.  
 {ces,ran}@cs.stir.ac.uk

**Abstract.** We introduce a series of descriptions of disease spread using the process algebra WSCCS and compare the derived mean field equations with the traditional ordinary differential equation model. Even the preliminary work presented here brings to light interesting theoretical questions about the “best” way to defined the model.



# ODE

Differentiating Processes!



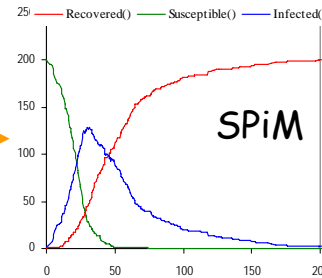
"useless" reactions

$$\begin{aligned}
 [S]^\bullet &= -t[S][I] \\
 [I]^\bullet &= t[S][I] - r[I] \\
 [R]^\bullet &= r[I]
 \end{aligned}$$

Automata produce the standard ODEs!

$$\begin{aligned}
 \frac{dS}{dt} &= -aIS \\
 \frac{dI}{dt} &= aIS - bI \\
 \frac{dR}{dt} &= bI
 \end{aligned}$$

(the Kermack-McKendrick, or SIR model)



```

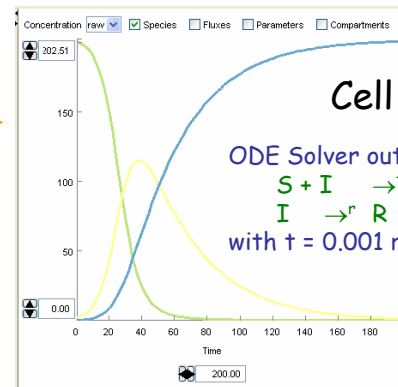
directive sample 500:0:1000
directive plot Recovered(), Susceptible(), Infected()

new infect @0.001:chan()
val recover = 0.03

let Recovered() =
  ?infect: Recovered()
and Susceptible() =
  ?infect: Infected()

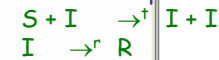
and Infected() =
  do !infect: Infected()
  or ?infect: Infected()
  or delay@recover: Recovered()

run (200 of Susceptible() | 2 of Infected())
    
```

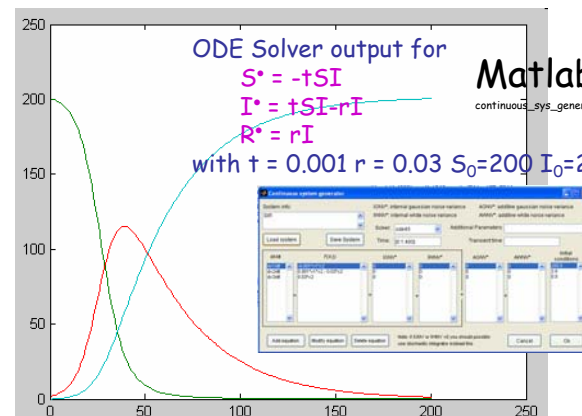


Cell Designer

ODE Solver output for reactions:



with  $t = 0.001$   $r = 0.03$   $[S]=200$   $[I]=2$



Matlab  
continuous\_sys\_generator

ODE Solver output for

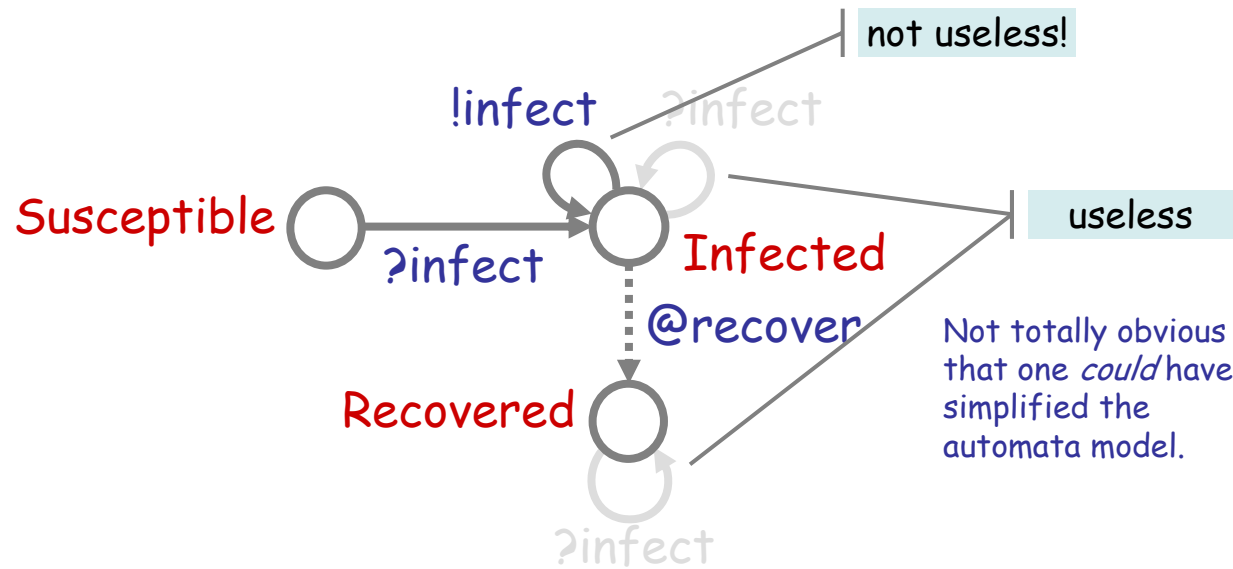
$$\begin{aligned}
 S^\bullet &= -tSI \\
 I^\bullet &= tSI - rI \\
 R^\bullet &= rI
 \end{aligned}$$

with  $t = 0.001$   $r = 0.03$   $S_0=200$   $I_0=2$

```

% ... (transcribed code from the right side of the slide) ...
    
```

# Simplified Model



```
directive sample 500.0 1000
directive plot Recovered(); Susceptible(); Infected()

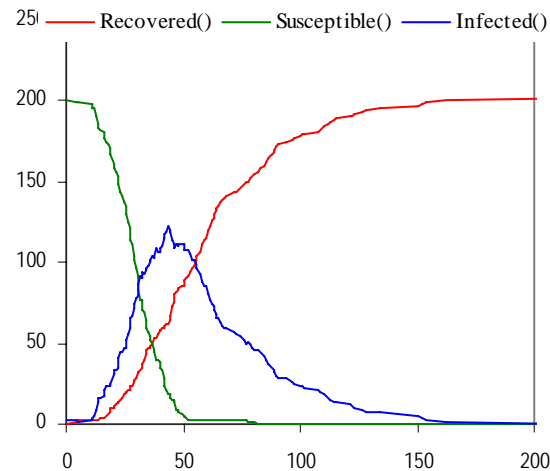
new infect @0.001:chan()
val recover = 0.03

let Recovered() =
  ()

and Susceptible() =
  ?infect; Infected()

and Infected() =
  do !infect; Infected()
  or delay@recover; Recovered()

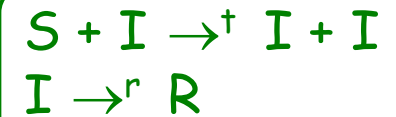
run (200 of Susceptible() | 2 of Infected())
```



$$S = ?i_{(t)}; I$$

$$I = !i_{(t)}; I \oplus \tau_r; R$$

$$R = 0$$



$$[S]' = -t[S][I]$$

$$[I]' = t[S][I] - r[I]$$

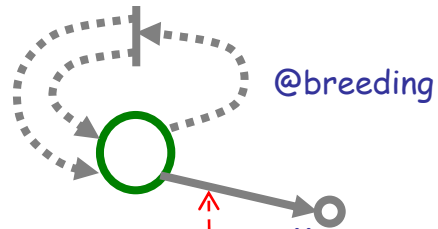
$$[R]' = r[I]$$

Same ODE, hence equivalent automata models.

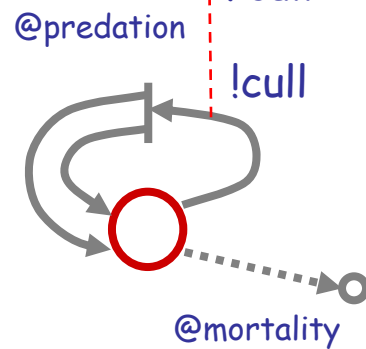
# Lotka-Volterra

# Predator-Prey

Herbivor



Carnivor



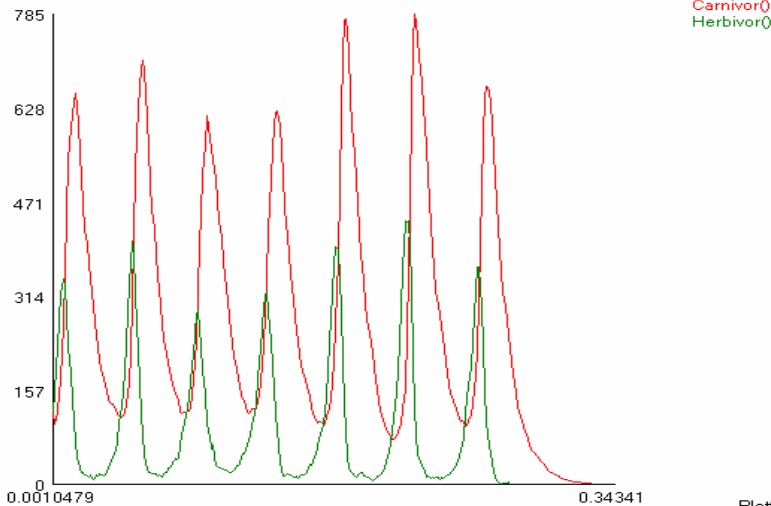
```
directive sample 1.0 1000
directive plot Carnivor(); Herbivor()
```

```
val mortality = 100.0
val breeding = 300.0
val predation = 1.0
new cull @predation:chan()
```

```
let Herbivor() =
  do delay@breeding; (Herbivor() | Herbivor())
  or ?cull; ()
```

```
and Carnivor() =
  do delay@mortality; ()
  or !cull; (Carnivor() | Carnivor())
```

```
run 100 of Herbivor()
run 100 of Carnivor()
```

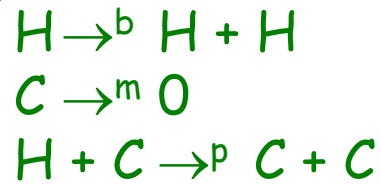
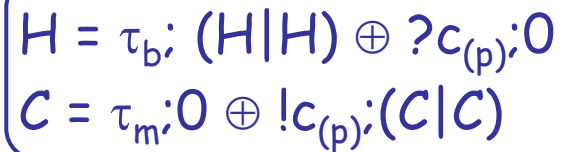


Simulation: Halted, Time = 0.343410 (317 points at 0.0068489 simTime/sysTime)

Plotting: Live

*An unbounded  
state system!*

# ODE



$$[H]' = b[H] - p[H][C]$$

$$[C]' = -m[C] + p[H][C]$$

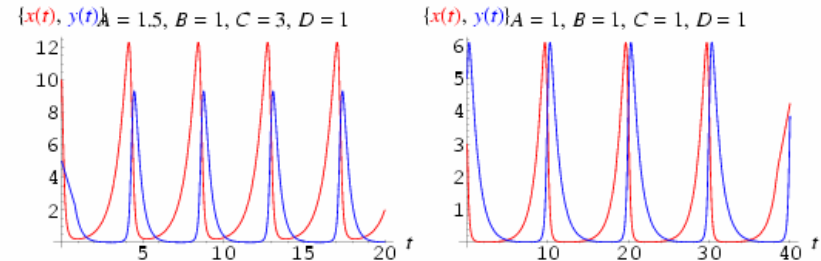
## Lotka-Volterra Equations



mathworld

The Lotka-Volterra equations describe an ecological predator-prey (or parasite-host) model which assumes that, for a set of fixed positive constants  $A$  (the growth rate of prey),  $B$  (the rate at which predators destroy prey),  $C$  (the death rate of predators), and  $D$  (the rate at which predators increase by consuming prey), the following conditions hold.

1. A prey population  $x$  increases at a rate  $dx = Ax dt$  (proportional to the number of prey) but is simultaneously destroyed by predators at a rate  $dx = -Bxy dt$  (proportional to the product of the numbers of prey and predators).
2. A predator population  $y$  decreases at a rate  $dy = -Cy dt$  (proportional to the number of predators), but increases at a rate  $dy = Dxy dt$  (again proportional to the product of the numbers of prey and predators).



This gives the coupled differential equations

$$\frac{dx}{dt} = Ax - Bxy \quad (1)$$

$$\frac{dy}{dt} = -Cy + Dxy, \quad (2)$$

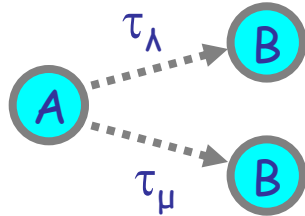
Automata produce the Lotka-Volterra model (with  $B=D$ )



# Laws by ODEs

# Choice Law by ODEs

$$\tau_\lambda;B \oplus \tau_\mu;B = \tau_{\lambda+\mu};B$$



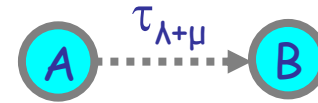
$$A = \tau_\lambda;B \oplus \tau_\mu;B$$



$$\begin{array}{l} A \xrightarrow{\lambda} B \\ A \xrightarrow{\mu} B \end{array}$$



$$\begin{array}{l} [A]^\bullet = -\lambda[A] - \mu[A] \\ [B]^\bullet = \lambda[A] + \mu[A] \end{array}$$



$$A = \tau_{\lambda+\mu};B$$



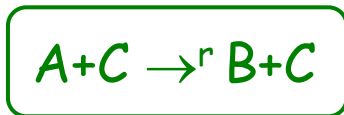
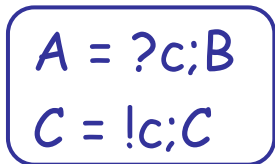
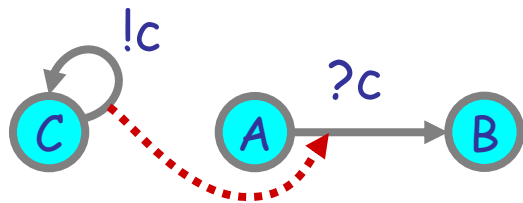
$$A \xrightarrow{\lambda+\mu} B$$



$$\begin{array}{l} [A]^\bullet = -(\lambda+\mu)[A] \\ [B]^\bullet = (\lambda+\mu)[A] \end{array}$$

=

# Idle Interaction Law by ODEs



$$[A]^\bullet = -r[A][C]$$

$$[B]^\bullet = r[A][C]$$

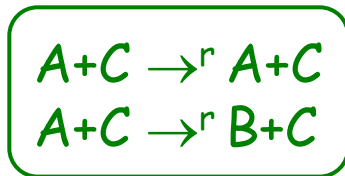
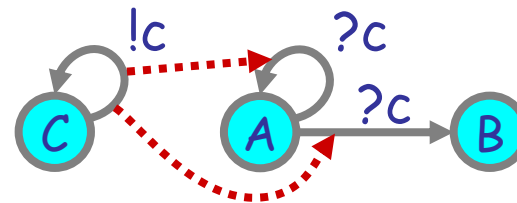
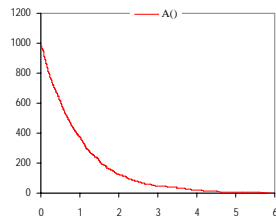
$$[C]^\bullet = 0$$

directive sample 6.0 1000  
directive plot A()

new c@1.0:chan

let A() = ?c; B()  
and B() = ()  
and C() = !c; C()

run (C) | 1000 of A()



$$[A]^\bullet = -r[A][C]$$

$$[B]^\bullet = r[A][C]$$

$$[C]^\bullet = 0$$

It may seem like A should decrease half as fast, but NO! Two ways to explain:

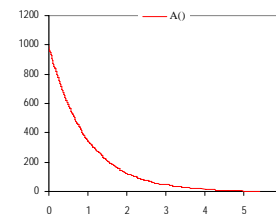
- State A is *memoryless* of any past idling.
- Activity on c is double

directive sample 6.0 1000  
directive plot A()

new c@1.0:chan

let A() = do ?c; B() or ?c; A()  
and B() = ()  
and C() = !c; C()

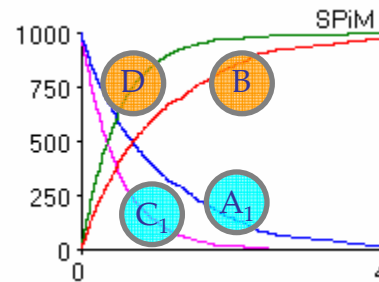
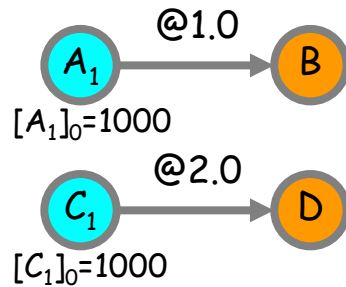
run (C) | 1000 of A()



# Stochastic Interleaving

$$\tau_\lambda;B \mid \tau_\mu;D = \tau_\lambda;(B \mid \tau_\mu;D) \oplus \tau_\mu;(\tau_\lambda;B \mid D)$$

Ex:  $\lambda=1.0, \mu=2.0$

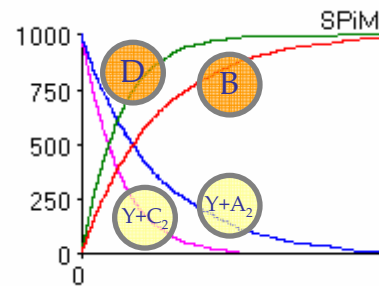
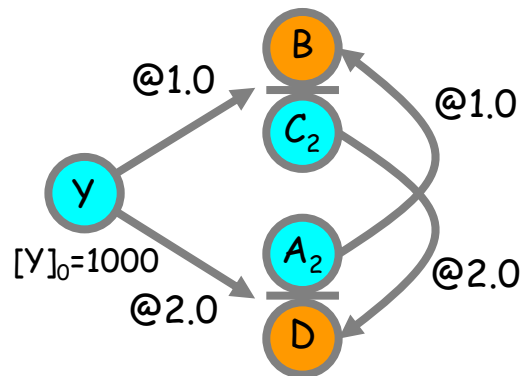


```
directive sample 4.0 10000
directive plot A(); B(); C(); D()
```

```
let A() = delay@1.0; B()
and B() = ()
```

```
let C() = delay@2.0; D()
and D() = ()
```

```
run 1000 of (A() | C())
```



```
directive sample 4.0 10000
directive plot
  ?YA; B(); ?YC; D(); Y(); A(); C()
new YA@1.0:chan new YC@1.0:chan
```

```
let A() = do delay@1.0; B() or ?YA
and B() = ()
```

```
let C() = do delay@2.0; D() or ?YC
and D() = ()
```

```
let Y() =
  do delay@1.0; (B() | C())
  or delay@2.0; (A() | D())
  or ?YA or ?YC
```

```
run 1000 of Y()
```

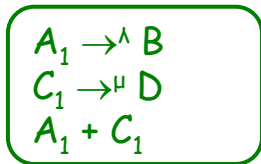
Amazingly, the B's and the D's from the two branches sum up to exponential distributions

# Stochastic Interleaving Law by ODEs

$$\tau_{\lambda};B \mid \tau_{\mu};D = \tau_{\lambda};(B \mid \tau_{\mu};D) \oplus \tau_{\mu};(\tau_{\lambda};B \mid D)$$

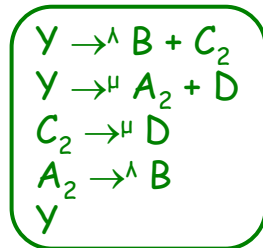
Want to show that B and D on both sides have the "same behavior" (equal quantities of B and D produced at all times)

$$\begin{aligned} A_1 &= \tau_{\lambda};B \\ C_1 &= \tau_{\mu};D \\ A_1 \mid C_1 \end{aligned}$$



$$\begin{aligned} [A_1]^{\bullet} &= -\lambda[A_1] \\ [B]^{\bullet} &= \lambda[A_1] \\ [C_1]^{\bullet} &= -\mu[C_1] \\ [D]^{\bullet} &= \mu[C_1] \end{aligned}$$

$$\begin{aligned} Y &= \tau_{\lambda};(B \mid C_2) \oplus \tau_{\mu};(A_2 \mid D) \\ C_2 &= \tau_{\mu};D \\ A_2 &= \tau_{\lambda};B \\ Y \end{aligned}$$



$$\begin{aligned} [Y]^{\bullet} &= -\lambda[Y] - \mu[Y] \\ [A_2]^{\bullet} &= \mu[Y] - \lambda[A_2] \\ [B]^{\bullet} &= \lambda[Y] + \lambda[A_2] \\ [C_2]^{\bullet} &= \lambda[Y] - \mu[C_2] \\ [D]^{\bullet} &= \mu[Y] + \mu[C_2] \end{aligned}$$

=?

$$\begin{aligned} [Y+A_2]^{\bullet} &= -\lambda[Y+A_2] \\ [B]^{\bullet} &= \lambda[Y+A_2] \\ [Y+C_2]^{\bullet} &= -\mu[Y+C_2] \\ [D]^{\bullet} &= \mu[Y+C_2] \end{aligned}$$

$$\begin{aligned} [Y+A_2]^{\bullet} &= [Y]^{\bullet} + [A_2]^{\bullet} \\ &= -\lambda[Y] - \mu[Y] + \mu[Y] - \lambda[A_2] \\ &= -\lambda[Y] - \lambda[A_2] \\ &= -\lambda[Y+A_2] \end{aligned} \quad [Y+A_2] \text{ decays exponentially!}$$

[B] and [D] have equal time evolutions on the two sides provided that  $[A_1]=[Y+A_2]$  and  $[C_1]=[Y+C_2]$ . This imposes the constraint, in particular, that  $[A_1]_0=[Y+A_2]_0$  and  $[C_1]_0=[Y+C_2]_0$  (at time zero). The initial conditions of the right hand system specify that  $[A_2]_0=[C_2]_0=0$  (since only Y is present). Therefore, we obtain that  $[A_1]_0=[C_1]_0=[Y]_0$ .

So, for example, if we run a stochastic simulation of the left hand side with  $1000 \cdot A_1$  and  $1000 \cdot C_1$ , we obtain the same curves for B and D than a stochastic simulation of the right hand side with  $1000 \cdot Y$ .

# Biochemistry

## Interaction+Complexation



# Bidirectional Polymerization

new c@μ new stop@1.0

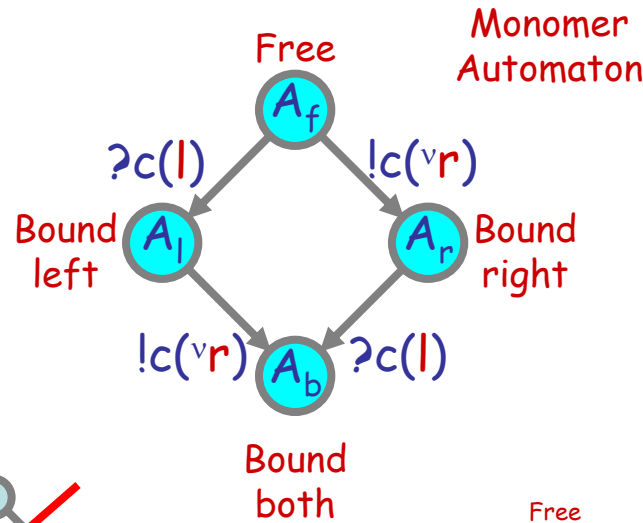
$A_{free} =$   
 $!c(\nu_{rht}, \lambda); A_{brht}(rht) +$   
 $?c(lft); A_{blft}(lft)$

$A_{blft}(lft) =$   
 $!c(\nu_{rht}, \lambda); A_{bound}(lft, rht)$

$A_{brht}(rht) =$   
 $?c(lft); A_{bound}(lft, rht)$

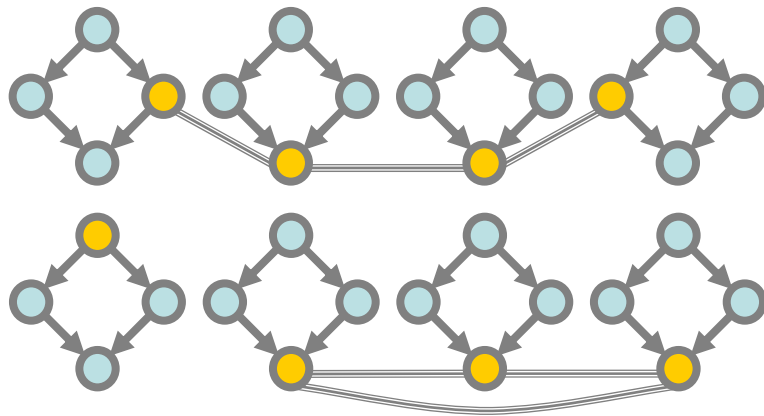
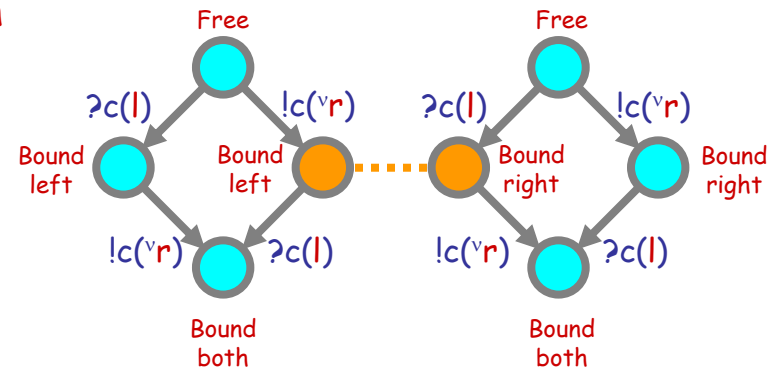
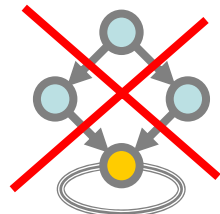
$A_{bound}(lft, rht) = ?stop$

Polymerization is iterated complexation.



## Polyautomata

Bound output  $!c(\nu_r)$  and input  $?c(l)$  on automata transitions to model complexation



```

directive sample 10000 0
directive plot AFree[] ABlft[] ABrt[] ABound[]

val lam = 1.0 val mu = 1.0
new c@μchan(chan) new stop@1.0chan

let AFree() =
  (new rht@lamchan run
   !c(νr); ABrt(rht))
  or ?c(lft); ABlft(lft)

and ABlft(lftchan) =
  (new rht@lamchan run
   !c(νr); ABound(lft,rht))

and ABrt(rhtchan) =
  ?c(lft); ABound(lft,rht)

and ABound(lftchan, rhtchan) =
  ?stop

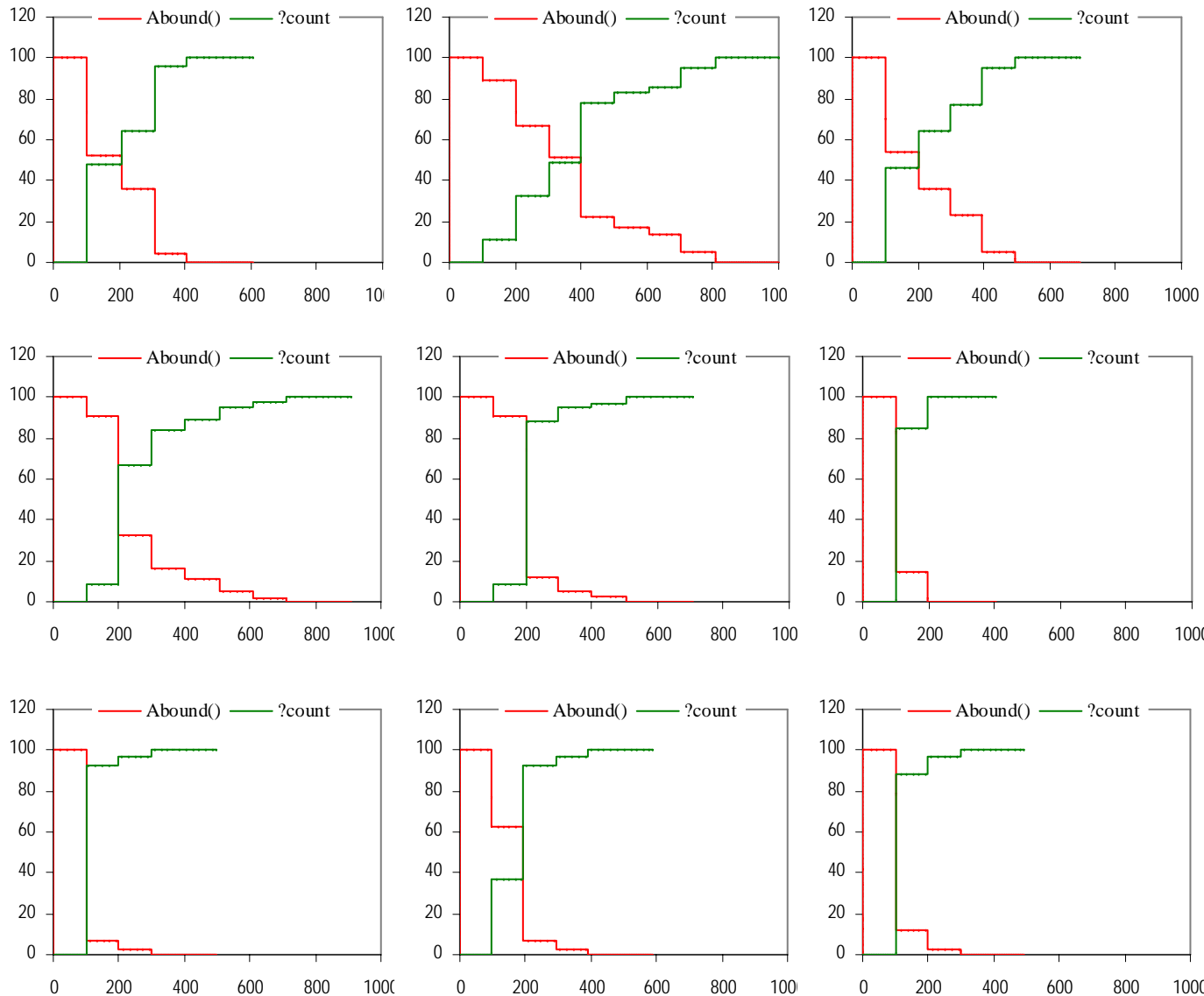
run (2 of AFree())
  
```

# Bidirectional Polymerization

## Circular Polymer Lengths

Scanning and counting the size of the circular polymers (by a cheap trick).

Polymer formation is complete within 10t; then a different polymer is scanned every 100t.



```
directive sample 1000.0
directive plot Abound(); ?count

type Link = chan(chan)
type Barb = chan

val lam = 1000.0 (* set high for better counting *)
val mu = 1.0
new c@mu:chan(Link)
new enter@lam:chan(Barb)
new count@lam:Barb

let Afree() =
  (new rht@lam:Link run
   do !c(rht); Abrht(rht)
   or ?c(lft); Ablft(lft))

and Ablft(lft:Link) =
  (new rht@lam:Link run
   !c(rht); Abound(lft,rht))

and Abrht(rht:Link) =
  ?c(lft); Abound(lft,rht)

and Abound(lft:Link, rht:Link) =
  do ?enter(barb); (?barb | !rht(barb))
  or ?lft(barb); (?barb | !rht(barb))
(* each Abound waits for a barb, exhibits it, and passes it to
the right so we can plot number of Abound in a ring *)

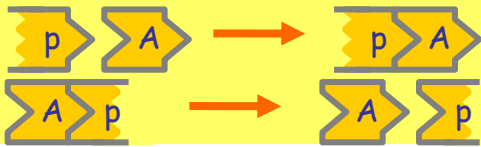
let clock(t:float, tick:chan) = (* sends a tick every t time *)
  (val ti = t/1000.0 val d = 1.0/ti
   let step(n:int) =
     if n<=0 then !tick; clock(t,tick) else delay@d; step(n-1)
   run step(1000))

new tick:chan
let Scan() = ?tick; !enter(count); Scan()

run 100 of Afree()
run (clock(100.0, tick) | Scan())
```

$100 \times A_{free}$ , initially.  
 The height of each rising step is the size of a separate circular polymer. (Unbiased sample of nine consecutive runs.)





# Actin-like Poly/Depolymerization

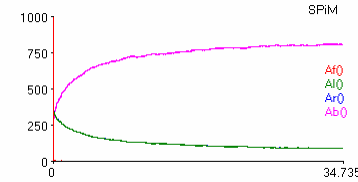
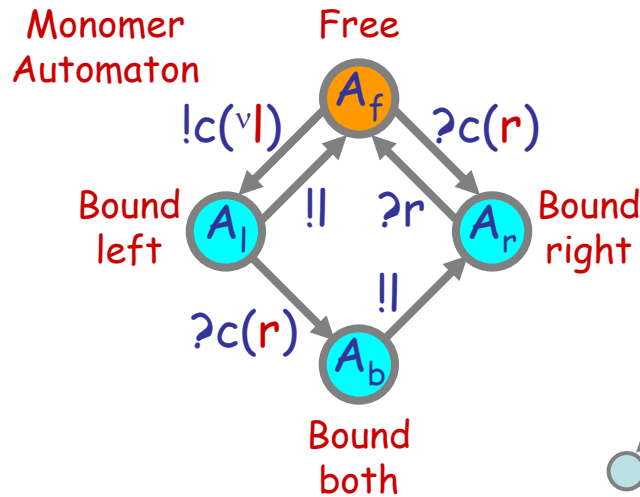
new c@μ

$$A_{free} = !c(v|); A_{blft}(lft) + ?c(rht); A_{brht}(rht)$$

$$A_{blft}(lft) = !lft; A_{free} + ?c(rht); A_{bound}(lft,rht)$$

$$A_{brht}(rht) = ?rht; A_{free}$$

$$A_{bound}(lft,rht) = !lft; A_{brht}(rht)$$



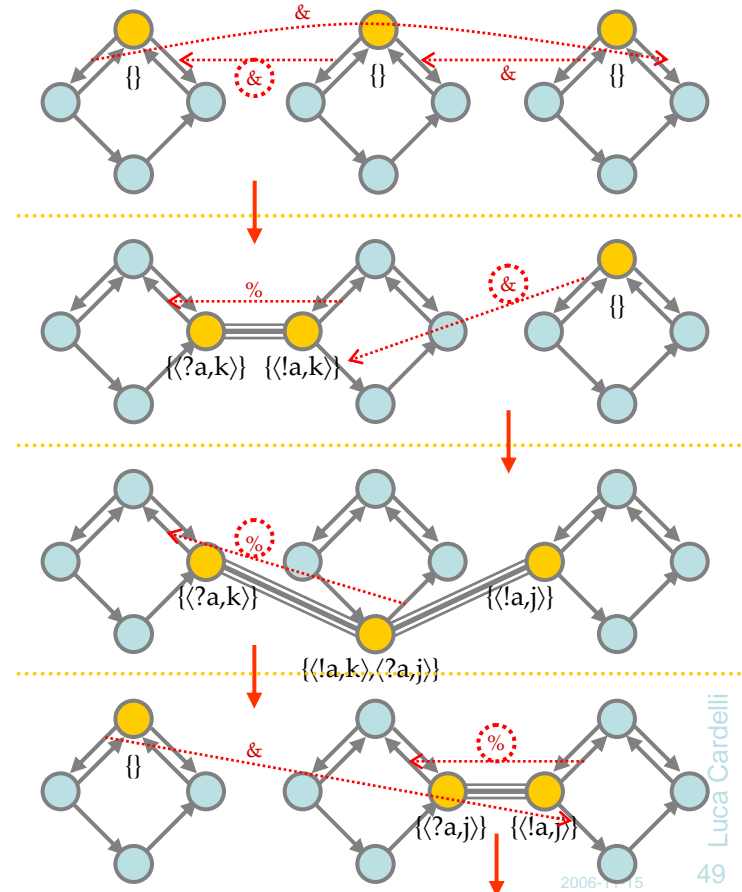
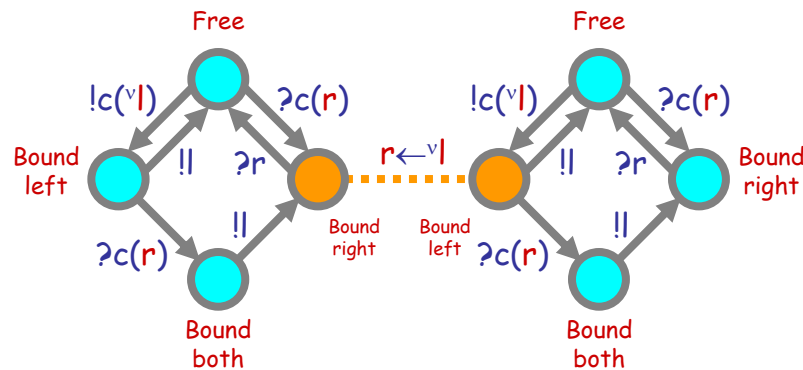
```

directive sample 1000.0
directive plot A_f, A_l, A_r, A_b

val sim = 1.0 ("disoc")
val mu = 1.0 ("asse")
new c@mu:chan(chan)

let A_f() =
  (new lft@l:chan run
   do !c(lft), A_l(lft)
   or ?c(rht), A_r(rht))
  and A_l(lft:chan) =
    do lft, A_f()
    or ?c(rht), A_b(lft,rht)
  and A_r(rht:chan) =
    ?rht, A_f()
  and A_b(lft:chan, rht:chan) =
    !lft, A_r(rht)
run 1000 of A_f()
  
```

1000 monomers settle to  
~100 polymers of size ~10



# Parametric Processes

# Chemical Parametric Form (CPF)

$E ::= X_1(\mathbf{p}_1)=M_1, \dots, X_n(\mathbf{p}_n)=M_n$

$M ::= \pi_1:P_1 \oplus \dots \oplus \pi_n:P_n$

$P ::= X_1(\mathbf{p}_1) \mid \dots \mid X_n(\mathbf{p}_n)$

$\pi ::= \tau_r \ ?n(\mathbf{p}) \ !n(\mathbf{p})$

$CPF ::= E, P$

Definitions  $(n \geq 0)$

Molecules  $(n \geq 0)$

Solutions  $(n \geq 0)$

Interactions

with initial conditions

Not bounded-state systems.

Not finite-control systems.

But still **finite-species** systems.

$\oplus$  is stochastic choice (vs. + for chemical reactions)

0 is the null solution ( $P|0 = 0|P = P$ )

and null molecule ( $M \oplus 0 = 0 \oplus M = M$ ) ( $\tau_0:P = 0$ )

$X_i$  are distinct in  $E$ ,  $\mathbf{p}$  are vectors of names

$\mathbf{p}$  are vectors of distinct names when in **binding position**

Each free name  $n$  in  $E$  is assigned a fixed rate  $r$ :

written either  $n_{(r)}$ , or  $\rho_{CPF}(n)=r$ .

A translation from CPF to CGF exists

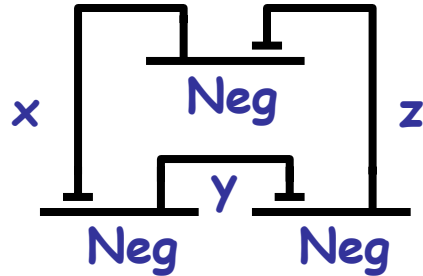
(expanding all possible instantiation of parameters from the initial conditions)

An incremental translation algorithm exists

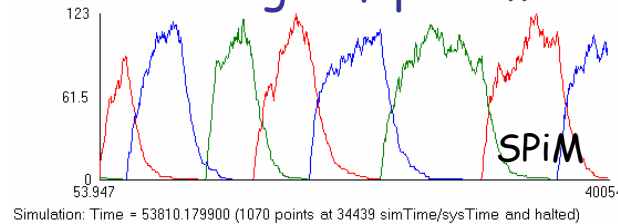
(expanding on demand from initial conditions)

# And Yet It Moves

## The Repressilator



A fine stochastic oscillator over a wide range of parameters.



```

directive sample 50000.0 1000
directive plot lc: lb: lc

val dk = 0.001 (* Decay rate *)
val inh = 0.001 (* Inhibition rate *)
val cst = 0.1 (* Constitutive rate *)

let tr(p:chan()) = do lp: trfp() or delay@dk

let neg(a:chan(), b:chan()) =
  do do: delay@inh; neg(a,b)
  or delay@cst; tr(b) | neg(a,b)

val bind = 1.0 (* Protein binding rate *)
new a@bind;chan() new b@bind;chan() new
c@bind;chan()
run (neg(c,a) | neg(a,b) | neg(b,z))
    
```

## Parametric representation

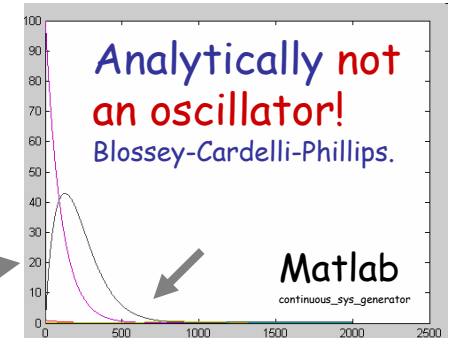
$$\begin{aligned}
 \text{Neg}(a,b) &= ?a; \text{Inh}(a,b) \oplus \tau_{\varepsilon}; (\text{Tr}(b) | \text{Neg}(a,b)) \\
 \text{Inh}(a,b) &= \tau_{\eta}; \text{Neg}(a,b) \\
 \text{Tr}(b) &= !b; \text{Tr}(b) \oplus \tau_{\gamma}; 0 \\
 \text{Neg}(x_{(r)}, y_{(r)}) &| \text{Neg}(y_{(r)}, z_{(r)}) | \text{Neg}(z_{(r)}, x_{(r)})
 \end{aligned}$$

$$\begin{aligned}
 [\text{Neg}/x,y]^* &= -r[\text{Tr}/x][\text{Neg}/x,y] + \eta[\text{Inh}/x,y] \\
 [\text{Neg}/y,z]^* &= -r[\text{Tr}/y][\text{Neg}/y,z] + \eta[\text{Inh}/y,z] \\
 [\text{Neg}/z,x]^* &= -r[\text{Tr}/z][\text{Neg}/z,x] + \eta[\text{Inh}/z,x] \\
 [\text{Inh}/x,y]^* &= r[\text{Tr}/x][\text{Neg}/x,y] - \eta[\text{Inh}/x,y] \\
 [\text{Inh}/y,z]^* &= r[\text{Tr}/y][\text{Neg}/y,z] - \eta[\text{Inh}/y,z] \\
 [\text{Inh}/z,x]^* &= r[\text{Tr}/z][\text{Neg}/z,x] - \eta[\text{Inh}/z,x] \\
 [\text{Tr}/x]^* &= \varepsilon[\text{Neg}/z,x] - \gamma[\text{Tr}/x] \\
 [\text{Tr}/y]^* &= \varepsilon[\text{Neg}/x,y] - \gamma[\text{Tr}/y] \\
 [\text{Tr}/z]^* &= \varepsilon[\text{Neg}/y,z] - \gamma[\text{Tr}/z]
 \end{aligned}$$

$$\begin{aligned}
 \text{Neg}/x,y &\rightarrow^{\varepsilon} \text{Tr}/y + \text{Neg}/x,y \\
 \text{Neg}/y,z &\rightarrow^{\varepsilon} \text{Tr}/z + \text{Neg}/y,z \\
 \text{Neg}/z,x &\rightarrow^{\varepsilon} \text{Tr}/x + \text{Neg}/z,x \\
 \text{Tr}/x + \text{Neg}/x,y &\rightarrow^r \text{Tr}/x + \text{Inh}/x,y \\
 \text{Tr}/y + \text{Neg}/y,z &\rightarrow^r \text{Tr}/y + \text{Inh}/y,z \\
 \text{Tr}/z + \text{Neg}/z,x &\rightarrow^r \text{Tr}/z + \text{Inh}/z,x \\
 \text{Inh}/x,y &\rightarrow^{\eta} \text{Neg}/x,y \\
 \text{Inh}/y,z &\rightarrow^{\eta} \text{Neg}/y,z \\
 \text{Inh}/z,x &\rightarrow^{\eta} \text{Neg}/z,x \\
 \text{Tr}/x &\rightarrow^{\gamma} 0 \\
 \text{Tr}/y &\rightarrow^{\gamma} 0 \\
 \text{Tr}/z &\rightarrow^{\gamma} 0 \\
 \text{Neg}/x,y + \text{Neg}/y,z + \text{Neg}/z,x &
 \end{aligned}$$

simplifying (N is the quantity of each of the 3 gates)

$$\begin{aligned}
 [\text{Neg}/x,y]^* &= \eta N - (\eta + r[\text{Tr}/x])[\text{Neg}/x,y] \\
 [\text{Neg}/y,z]^* &= \eta N - (\eta + r[\text{Tr}/y])[\text{Neg}/y,z] \\
 [\text{Neg}/z,x]^* &= \eta N - (\eta + r[\text{Tr}/z])[\text{Neg}/z,x] \\
 [\text{Tr}/x]^* &= \varepsilon[\text{Neg}/z,x] - \gamma[\text{Tr}/x] \\
 [\text{Tr}/y]^* &= \varepsilon[\text{Neg}/x,y] - \gamma[\text{Tr}/y] \\
 [\text{Tr}/z]^* &= \varepsilon[\text{Neg}/y,z] - \gamma[\text{Tr}/z]
 \end{aligned}$$



```

interval/step (0:10:20000)    N=1, r=1.0, \varepsilon=0.1, \eta=0.001, \gamma=0.001
(Neg/x,y)    dx1/dt = 0.001 - (0.001 + x4)*x1    1.0
(Neg/y,z)    dx2/dt = 0.001 - (0.001 + x5)*x2    1.0
(Neg/z,x)    dx3/dt = 0.001 - (0.001 + x6)*x3    1.0
(Tr/x)       dx4/dt = 0.1*x3 - 0.001*x4          100.0
(Tr/y)       dx5/dt = 0.1*x1 - 0.001*x5          0
(Tr/z)       dx6/dt = 0.1*x2 - 0.001*x6          0
    
```

# Conclusions

# Conclusions

- **Compositional Models**
  - Accurate (at the "appropriate" abstraction level).
  - Manageable (so we can scale them up by composition).
- **Interacting Automata**
  - Complex global behavior from simple components.
  - Bridging individual and collective behavior.
  - Connections to classical Markov theory, chemical Master Equation, and Rate Equation.
- **Mapping out "the whole system"**
  - Through an "artificial biochemistry" (a scalable mathematical and computational modeling framework) to investigate "real biochemistry" on a large scale.

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